

Research Paper

Factors associated with death outcome in patients with severe coronavirus disease-19 (COVID-19): a case-control study

Feng Pan^{*1,2}; Lian Yang^{*1,2}; Yuncheng Li^{*3}; Bo Liang^{1,2}; Lin Li^{1,2}; Tianhe Ye^{1,2}; Lingli Li^{1,2}; Dehan Liu^{1,2}; Shan Gui^{1,2}; Yu Hu⁴; Chuansheng Zheng^{1,2}✉

1. Department of Radiology, Union Hospital, Tongji Medical College, Huazhong University of Science and Technology, Jiefang Avenue #1277, Wuhan, 430022, China.
2. Hubei Province Key Laboratory of Molecular Imaging, Wuhan, 430022, China.
3. Department of Otorhinolaryngology, Union Hospital, Tongji Medical College, Huazhong University of Science and Technology, 1277 Jiefang Avenue, Wuhan 430022, China.
4. Institute of Hematology, Union Hospital, Tongji Medical College, Huazhong University of Science and Technology, 1277 Jiefang Avenue, Wuhan 430022, China.

*These three authors contributed equally to this manuscript.

✉ Corresponding author: Chuansheng Zheng, Prof, MD, PhD, Chief in Department of Radiology. Email: hqzcsxh@sina.com; Phone: +86 027 85353238.

¹Department of Radiology, Union Hospital, Tongji Medical College, Huazhong University of Science and Technology, ²Hubei Province Key Laboratory of Molecular Imaging, No. 1277 Jiefang Avenue, Wuhan, China. Yu Hu, Prof, MD, PhD. Email: dr_huyu@126.com; Phone: +86 27 85726387; Fax: +86 27 85726387.

⁴Institute of Hematology, Union Hospital, Tongji Medical College, Huazhong University of Science and Technology, No. 1277 Jiefang Avenue, Wuhan, China

© The author(s). This is an open access article distributed under the terms of the Creative Commons Attribution License (<https://creativecommons.org/licenses/by/4.0/>). See <http://ivyspring.com/terms> for full terms and conditions.

Received: 2020.04.09; Accepted: 2020.05.05; Published: 2020.05.18

Abstract

Rationale: Up to date, the exploration of clinical features in severe COVID-19 patients were mostly from the same center in Wuhan, China. The clinical data in other centers is limited. This study aims to explore the feasible parameters which could be used in clinical practice to predict the prognosis in hospitalized patients with severe coronavirus disease-19 (COVID-19).

Methods: In this case-control study, patients with severe COVID-19 in this newly established isolation center on admission between 27 January 2020 to 19 March 2020 were divided to discharge group and death event group. Clinical information was collected and analyzed for the following objectives: 1. Comparisons of basic characteristics between two groups; 2. Risk factors for death on admission using logistic regression; 3. Dynamic changes of radiographic and laboratory parameters between two groups in the course.

Results: 124 patients with severe COVID-19 on admission were included and divided into discharge group (n=35) and death event group (n=89). Sex, SpO₂, breath rate, diastolic pressure, neutrophil, lymphocyte, C-reactive protein (CRP), procalcitonin (PCT), lactate dehydrogenase (LDH), and D-dimer were significantly correlated with death events identified using bivariate logistic regression. Further multivariate logistic regression demonstrated a significant model fitting with C-index of 0.845 (p<0.001), in which SpO₂≤89%, lymphocyte≤0.64×10⁹/L, CRP>77.35mg/L, PCT>0.20μg/L, and LDH>481U/L were the independent risk factors with the ORs of 2.959, 4.015, 2.852, 3.554, and 3.185, respectively (p<0.04). In the course, persistently lower lymphocyte with higher levels of CRP, PCT, IL-6, neutrophil, LDH, D-dimer, cardiac troponin I (cTnI), brain natriuretic peptide (BNP), and increased CD4+/CD8+ T-lymphocyte ratio and were observed in death events group, while these parameters stayed stable or improved in discharge group.

Conclusions: On admission, the levels of SpO₂, lymphocyte, CRP, PCT, and LDH could predict the prognosis of severe COVID-19 patients. Systematic inflammation with induced cardiac dysfunction was likely a primary reason for death events in severe COVID-19 except for acute respiratory distress syndrome.

Key words: COVID-19; Critical care; Prognosis; Risk factors; Radiography

Introduction

Since the outbreak of COVID-19 in Wuhan, it spread rapidly worldwide with a severity spectrum from asymptomatic to critical illness with the overall rate of approximately 3% [1-3]. Up to date, an increasing number of more than forty-thousand fatalities were reported [4]. Owing to the genomic homologies of the pathogen from the coronaviruses causing the Severe Acute Respiratory Syndrome (SARS) and the Middle East Respiratory Syndrome (MERS), COVID-19 presented a similar clinical course and pathological feature [5-8]. Although most of the COVID-19 patients were mild who could gradually recover after two weeks, about 15-20% of patients developed severe interstitial pneumonia [1, 8, 9]. Patients with severe illness usually developed acute respiratory distress syndrome (ARDS) that requires invasive mechanical ventilation therapy in the intensive care unit (ICU), and the mortality rate was reported around 50-60% [1, 10].

To date, the clinical studies revealed different clinical and laboratory features between survivors and non-survivors, or between the non-severe and severe patients, and some prognostic risk factors were preliminarily discovered, such as age and D-dimer [6-8, 10-13]. However, most of these results were from or involving the same center in Wuhan, China, at a similar period, leading to potential selection bias. Moreover, the same patients in different articles were suspected [14]. This study systematically analyzed the clinical features, laboratory investigations, and radiographic estimation in a temporary isolation center in Wuhan, China, established specially for severe COVID-19 patients by an emergency directive of the Wuhan Anti-epidemic Headquarters since 27 January 2020. All severe patients had to fulfill the WHO criteria for admission to hospital for a respiratory illness, which was slightly modified and adopted by both the WHO and the Chinese National Diagnosis and Treatment Protocols to define a severe case of COVID-19 [15-18]. These criteria defined severe COVID-19 as fever and/or respiratory infection with positive COVID-19 nucleic acid detection, plus one of the following: respiratory rate > 30 breaths/min; severe respiratory distress; or SpO₂ ≤ 93% on room air. This study examined the factors associated with mortality in patients who fulfilled these criteria of severity at the time of hospital admission. It excluded patients who at the time of admission had, according to WHO criteria, mild illness, pneumonia, or had already progressed to ARDS, sepsis, or septic shock [17]. For further speculation, it tried to explore the fatal course of COVID-19.

Methods

This study was approved by the Ethics of Committees of Union Hospital, Tongji Medical College, Huazhong University of Science and Technology (No. 2020-0026), and followed the 1964 Helsinki Declaration and its later amendments or comparable ethical standards. Only the anonymous data was collected and informed consent for this retrospective study was waived.

Patients

The consecutive admission records were reviewed retrospectively for the period from 27 January 2020 to 19 March 2020 in this provisional center (Western Campus, Union Hospital, Tongji Medical College, Huazhong University of Science and Technology). Clinical data, including initial symptoms, past medical history, hospitalization date, treatment, and hematological and biochemical blood test results were obtained directly through the institutional electronic patient database. The inclusion criteria included: 1. Confirmed severe COVID-19 on admission as mentioned above [15-18]; 2. No comorbidities which might impair the immune function, such as autoimmune disease, hematological malignancies, recent chemotherapy, etc.; 3. With definite discharge or COVID-19 related death outcome.

Imaging Estimation

Regular bedside chest digital radiographs (DRs) were regularly performed every 1 to 3 days instead of conventional computed tomography (CT) scans after admission owing to the risk during the transfer of severe patients. A conventional semi-quantitative scoring system for DR was used to evaluate the pulmonary involvement area of the abnormalities including air-space and reticular opacities [19, 20]. Each lung was divided into three equidistant zones (upper, middle, and lower), resulting in a total of six zones to be scored. Each zone was visually scored from 0 to 4 as: score 0, 0% involvement; score 1, 1%-25% involvement; score 2, 26%-50% involvement; score 3, 51-75% involvement; and score 4, ≥76% involvement. The DR score was the sum of the score of each zone. The analysis was performed by two radiologists (CZ and LY, who had 26 and 22 years of experience in thoracic radiology, respectively) and the decisions were reached in consensus. All radiologists were blinded to the clinical progress of the patients to avoid information bias.

Groups and Study Goals

Two groups were divided based on the clinical outcomes: discharge group and death event group.

Three objectives were explored in this study: 1. Comparisons of basic characteristics between two groups, including the occurrence of ARDS, which was defined as worsening respiratory symptoms with $\text{PaO}_2/\text{FiO}_2 \leq 300$ mmHg based on the Berlin definition and WHO criteria [21, 22]; 2. Risk factors for death on admission; 3. Dynamic changes in DR scores and laboratory investigations between two groups in the follow-up.

Statistical Analysis

Statistical analysis was performed using IBM SPSS Statistics Software (version 24; IBM, New York, USA). Quantitative data were presented as median with inter-quartile range (IQR) and counting data were presented as the percentage of the total unless otherwise specified. The comparisons of the quantitative data were statistically evaluated using the Mann-Whitney U test, according to the non-normal distribution assessed by the Shapiro-Wilk test. The quantitative parameters were stratified into binary variables with the cutoffs of median values. The comparisons of counting data were evaluated using the Chi-square test. The bivariate and multivariate logistic regressions were used to investigate the risk factors for death events involving the stratified clinical, radiographic, and laboratory parameters with significant differences between two groups on admission, and the odds ratio (OR) with 95% confidence interval (95%CI) were calculated. Variables were excluded in the multivariate logistic regression if: 1. The univariate analysis demonstrated no statistical significance ($p < 0.05$); 2. The multivariate analysis demonstrated the probability of $p \geq 0.10$ when using forward-LR method; 3. Variable had obvious collinearity with other variables resulting in variance inflation factor (VIF) of more than 5 [23]. C-index of predicted probability calculated using the optimal multivariate logistic model was evaluated by ROC curve estimation. A p-value of < 0.05 was defined as the statistical significance.

Results

Basic characteristics and comparisons between two groups

After retrieving the electronic records, 124 patients were eventually included in the further analysis from 211 consecutive records (Figure 1). The basic information of included patients was summarized in Table 1. The median age of patients was 68 yrs (IQR 61-75 yrs), and 68.5% of patients were male. Half of the patients had hypertension history and 11 patients had COPD (GOLD II grade) history which was controlled well after quitting smoking and

ipratropium treatment. Fever and cough were the most common initial symptoms (85.5% and 69.4%, respectively). The median interval between the admission and symptom onset was 11 days (IQR 7-15 days). On admission, the median body temperature on admission reached 38.3 °C (IQR: 37.0-39.0°C). The median breath rate was 30/min (IQR: 23-33/min) and median SpO₂ was 89% (IQR: 82-92%). In most patients (123/124, 99.2%), DRs demonstrated bilateral opacities with the median DR score of 13 (IQR: 7-16). Multiple abnormalities of biochemical and hematological parameters were observed as well, including neutrophil, lymphocyte, C-reactive protein (CRP), procalcitonin (PCT), lactate dehydrogenase (LDH), D-dimer, cardiac troponin I (cTnI), and brain natriuretic peptide (BNP), and so on. In the course, 91 patients (73.4%) aggravated to ARDS with the median period of 7 days after admission (IQR 4-11 days), who were transferred to ICU afterwards. Eventually, 89 patients (71.8%) with ARDS complications died who were included in death event group, while 35 patients (28.2%) were recovered and discharged who were included in discharge group.

Compared between the two groups, the patients in death event group had significantly higher male proportion (75.3% vs. 51.4%), lower SpO₂ (85% vs. 90%), and higher breath rate (30 vs 25/min) (Table 2). In the course, ARDS was more predominantly occurred in death event group compared with discharge group (89 vs. 2). 10 patients with ARDS in death event group refused endotracheal intubation. The median survival period in death event group was 15 days (IQR 9-22 days), while the median hospitalized period in discharge group was 29 days (IQR 23-35 days) (Table 2). On admission, the bedside DRs showed larger pulmonary opacities in death event group compared with discharge group, but without a significant difference (14 vs. 11, $p = 0.289$) (Table 2). Besides, more abnormalities of biochemical and hematological parameters were observed in death event group compared with discharge group, such as neutrophil, lymphocyte, CRP, PCT, LDH, D-dimer, cTnI, and BNP (Table 2).

Risk factor estimation of death events

The bivariate logistic regression was performed on the clinical, radiographic, and laboratory parameters (Table 3). As a result, ten stratified parameters including male sex, $\text{SpO}_2 \leq 89\%$, breath rate $> 30/\text{min}$, diastolic pressure $\leq 80\text{mmHg}$, neutrophil $> 6.46 \times 10^9/\text{L}$, lymphocyte $\leq 0.64 \times 10^9/\text{L}$, CRP $> 77.35\text{mg/L}$, PCT $> 0.20\mu\text{g/L}$, LDH $> 481\text{U/L}$, and D-dimer $> 3.06\text{mg/L}$ showed significant risks with death events (Table 3). No obvious multicollinearity was observed with the variance inflation factor (VIF) value of less

than 1.6 in each involved variable (**Table S1**). Further multivariate logistic regression showed a significant model fitting involving stratified SpO₂, lymphocyte, CRP, PCT, and LDH. The results demonstrated that SpO₂≤89%, lymphocyte≤0.64×10⁹/L, CRP>77.35 mg/L, PCT>0.20μg/L, and LDH>481U/L were the independent risk factors with the ORs of 2.959, 4.015, 2.852, 3.554, and 3.185, respectively (p<0.04). The C-index of the predicted probability calculated using this multivariate logistic model was 0.845 (p<0.001) (**Figure 2**).

Dynamic changes in laboratory investigations and DR scores between patients with clinical stability and death events

In the course, DR scores increased in death event group with a significant difference between the two groups after 1 to 3 days from admission (**Figure 3** and **4**). After 15 days, DR scores slowly declined in discharge group while increasing in death event group (**Figure 3** and **4**). In death event group, the neutrophil gradually increased to more than 10.00×10⁹/L, and lymphocyte stayed at the lower level from 0.47 to 0.56×10⁹/L in the course, with significant differences from discharge group (**Figure 4**). In discharge group, lymphocyte gradually increased from 0.88×10⁹/L to normal level (≥1.1×10⁹/L) after nine days. Besides, it demonstrated the significantly higher levels of CRP, PCT, LDH, and D-dimer in death event group, which gradually decreased or stayed at the relatively lower levels in

discharge group (**Figure 4**). cTnI and BNP demonstrated dramatic fluctuation with significantly higher levels at some time points in death event group compared with discharge group (**Figure 4**). No obvious abnormalities of total bilirubin (TB), aspartate aminotransferase (AST), and serum creatinine (Scr) in the course were observed in both groups (**Table S2**).

Among all 124 patients, 77 patients underwent consecutive cytokine profile and lymphocyte subsets analysis in the course, including 29 patients in charge group and 48 patients in death event group. The levels of IL-2, IL-6, TNF-α, and INF-γ didn't show significant differences between two groups and stayed in the normal reference ranges at most of time (**Figure 5**). Compared with discharge group, IL-6 was significantly elevated in death event group. And IL-10 was significantly increased after 29 days in death event group (**Figure 5**). CD3+, CD4+, and CD8+ T-lymphocyte ratio stayed in the normal range in both groups, though approaching the upper or lower reference at some time points (**Figure 6**). CD3+ and CD8+ T-lymphocyte ratios were significantly higher in discharge group at several time points, but the CD4+/CD8+ T-lymphocyte ratio seemed higher in death group compared with discharge group reaching significant difference at the time point of 17-19 days (**Figure 6**). The quantitative data were summarized in **Table S3**.

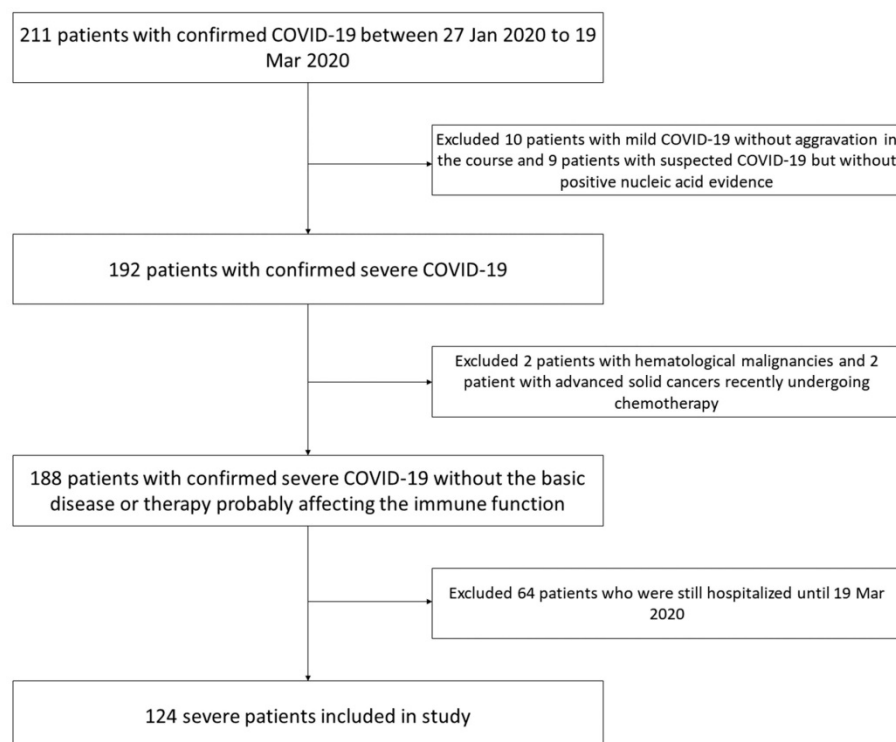


Figure 1. Flowchart of inclusion of the patients.

Table I. Baseline characteristics.

	Normal reference range	Total (n=124)
Age (y)		68 (61-75)
Sex		
Male		85 (68.5)
Female		39 (31.5)
Medical history		
Hypertension		62 (50.0)
Cardiovascular disease (CVD)		19 (15.3)
Diabetes mellitus		25 (20.2)
Chronic obstructive pulmonary disease (COPD)		11 (8.9)
Initial symptoms		
Fever		106 (85.5)
Low grade fever (37.5 - 38.0 °C)		24 (19.4)
Moderate fever (38.1 - 39.0 °C)		42 (33.9)
High-grade fever (≥39.1 °C)		40 (32.3)
Cough		86 (69.4)
Fatigue		61 (49.2)
Chest distress		57 (46.0)
Expectoration		49 (39.5)
Chills		27 (21.8)
Myalgia		26 (21.0)
Diarrhea		19 (15.3)
Time of admission after symptom onset (d)		11 (7-15)
Vital signs on admission		
SpO ₂ (%)		89 (82-92)
Body temperature (°C)		38.3 (37.0-39.0)
Heart rate (/min)		90 (80-102)
Breath rate (/min)		30 (23-33)
Systolic pressure (mmHg)		132 (122-150)
Diastolic pressure (mmHg)		80 (73-88)
Diagnosis of severe COVID-19		
Breath rate ≥30/min		6 (4.8)
SpO ₂ ≤93%		58 (46.8)
Both		60 (48.4)
DR estimations on admission		
Unilateral involvement		1 (0.8)
Bilateral involvement		123 (99.2)
DR score	(0)	13 (7-16)
Laboratory investigations on admission		
White blood cell (WBC) (×10 ⁹ /L)	(3.50-9.50)	7.52 (5.07-10.92)
Neutrophil (×10 ⁹ /L)	(1.80-6.30)	6.46 (3.91-9.92)
Lymphocyte (×10 ⁹ /L)	(1.10-3.20)	0.64 (0.46-1.02)
Hemoglobin (g/L)	(115-150)	130 (121-142)
Platelet (×10 ⁹ /L)	(125-350)	187 (150-230)
C-reactive protein (CRP) (mg/L)	(0.00-8.00)	77.35 (43.13-111.94)
Procalcitonin (PCT) (µg/L)	(<0.05)	0.20 (0.09-0.40)
Total bilirubin (TB) (µmol/L)	(3.0-20.0)	13.6 (9.7-20.0)
Alanine aminotransferase (ALT) (U/L)	(5-35)	37 (25-59)
Aspartate aminotransferase (AST) (U/L)	(8-40)	47 (31-69)
Lactate dehydrogenase (LDH) (U/L)	(109-245)	481 (360-597)
Albumin (g/L)	(33.0-55.0)	28.2 (26.0-30.9)
Globulin (g/L)	(20.0-35.0)	34.2 (31.0-38.0)
Serum creatinine (Scr) (µmol/L)	(41.0-81.0)	74.1 (62.2-99.7)
Na ⁺ (mmol/L)	(137.0-147.0)	138.3 (135.8-140.4)
K ⁺ (mmol/L)	(3.5-5.3)	3.87 (3.53-4.24)
Cl ⁻ (mmol/L)	(96-108)	102.3 (99.6-105.2)
D-dimer (mg/L)	(<0.5)	3.06 (0.62-8.00)
Activated partial thromboplastin time (APTT) (s)	(27.0-45.0)	38.0 (33.1-42.5)
Prothrombin time (PT) (s)	(11.0-16.0)	13.9 (13.0-14.9)
Fibrinogen (g/L)	(2.00-4.00)	4.58 (3.45-5.25)
Cardiac troponin I (cTnI) (µg/L)	(<26.2)	19.3 (8.4-96.4)
Brain natriuretic peptide (BNP) (ng/L)	(<100.0)	79.3 (30.4-164.5)
Occurrence of ARDS in the course		91 (73.4)
Period between admission and occurrence of ARDS (d)		7 (4-11)
Hospitalized period (d)		19 (11-27)

Abbreviations: CVD: cardiovascular disease; COPD: chronic obstructive pulmonary disease; DR: digital radiograph; WBC: white blood cell count; CRP: C-reactive protein; PCT: procalcitonin; TB: total bilirubin; ALT: alanine aminotransferase; AST: aspartate aminotransferase; LDH: lactate dehydrogenase;

Scr: serum creatinine; APTT: activated partial thromboplastin time; PT: prothrombin time; cTnI: cardiac troponin I; BNP: brain natriuretic peptide.

Discussion

This study reported the differences between the discharged and died patients with severe COVID-19 in the clinical course. On admission, SpO₂ ≤89%, lymphocyte ≤0.64 × 10⁹/L, CRP >77.35 mg/L, PCT >0.20 µg/L, and LDH >481 U/L were the independent risk factors associated with death events. Besides, in the clinical course, significantly higher levels of DR score, neutrophil, IL-6, D-dimer, cTnI, and BNP were also observed in death event group. Thus, the results indicated that severe systematic inflammation with cardiac impairment played an important role in the fatal process of COVID-19.

Corresponding to the previous reports, old patients with a median age of 68 yrs (IQR 61-75 yrs) presenting more coexisting illnesses such as hypertension and CVD were more likely to develop severe COVID-19 [6, 8]. Male patients in this study were more inclined to occur ARDS and death events, in agreement with a retrospective report with a cohort of 44,672 patients where male patients over 60 years of age with comorbidities had a higher death risk [1]. In the course, patients in death event group showed a significantly higher ratio of ARDS than discharge group (100.0% vs. 5.7%). It indicated the survival rate was very low if the patients aggravated to this critical status and further exploration of the treatment to prevent ARDS is imperative. Although no significant difference in the period between initial symptom onset and admission was found between two groups, attention still should be paid to early diagnosis and treatment because the mean period of 11 days from symptom onset to admission in this study was probably too late when the severe progression had occurred [9].

Because the DR demonstrated a good consistency to chest CT which was also capable of estimating the COVID-19 in the follow-up, it was consecutively performed in this cohort in order to reduce the transferring risk of the severe patients and X-ray exposure [24]. On admission, bilaterally diffuse pulmonary opacities in DRs were observed in most patients (99.5%), similar to the demonstrations in SARS and MERS [19, 20]. The higher DR scores in death event group on admission were presented with a lower SpO₂ level and a higher breath rate when compared with discharge group, though no significant difference was revealed. However, more rapid progression of pulmonary opacities with a significantly higher DR score was observed afterwards in death event group. Different from no absorption in death event group, the pulmonary

lesions were slowly absorbed after half a month from admission in discharge group. Due to the limited medical resources in this provisional center, lung ultrasound was not routinely performed. From the previous studies, lung ultrasound could also be performed at the bedside and has been used in diagnosing and monitoring pneumonia with higher sensitivity and specificity ($\geq 94\%$) than DR but without X-ray exposure [25, 26]. Thus, it might be a good alternative to conventional bedside DR in severe COVID-19 patients.

Table 2. Comparisons between two groups Initial radiological estimations and laboratory investigations on admission.

	Discharge group (n=35)	Death events group (n=89)	p-value
Age (y)			
≤68	65 (49-77)	69 (61-73)	0.285
>68	20 (57.1)	42 (47.2)	0.318
Sex			
Male	18 (51.4)	67 (75.3)	0.010
Female	17 (48.6)	22 (24.7)	
Medical history			
Hypertension	15 (42.9)	47 (52.8)	0.318
CVD	6 (17.1)	13 (14.6)	0.724
Diabetes mellitus	6 (17.1)	19 (21.3)	0.599
COPD	4 (11.4)	7 (7.9)	0.530
Initial symptoms			
Fever	31 (88.6)	75 (84.3)	0.540
Low grade fever (37.5 - 38.0 °C)	10 (28.6)	14 (15.7)	0.432
Moderate fever (38.1 - 39.0 °C)	11 (31.4)	31 (34.8)	
High-grade fever (≥ 39.1 °C)	10 (28.6)	30 (33.7)	
Cough	28 (80.0)	58 (65.2)	0.107
Fatigue	16 (45.7)	45 (50.6)	0.627
Chest distress	15 (42.9)	42 (47.2)	0.663
Expectoration	14 (40.0)	35 (39.3)	0.945
Chills	6 (17.1)	21 (23.6)	0.433
Myalgia	6 (17.1)	20 (22.5)	0.512
Diarrhea	2 (5.7)	17 (19.1)	0.063
Time of admission after symptom onset (d)	11 (9-15)	10 (7-15)	0.354
≤11	21 (60.0)	48 (53.9)	0.540
>11	14 (40.0)	41 (46.1)	
Vital signs after admission			
SpO2 (%)	90 (88-92)	85 (80-90)	0.001
≤89	9 (25.7)	53 (59.6)	0.001
>89	26 (74.3)	36 (40.4)	
Body temperature (°C)	38.0 (36.6-39.0)	38.4 (37.4-39.0)	0.237
≤38.3	19 (54.3)	44 (49.4)	0.627
>38.3	16 (45.7)	45 (50.6)	
Heart rate (/min)	87 (76-103)	92 (80-102)	0.467
≤90	21 (60.0)	42 (47.2)	0.199
>90	14 (40.0)	47 (52.8)	
Breath rate (/min)	25 (20-30)	30 (25-34)	0.001
≤30	30 (85.7)	54 (60.7)	0.007
>30	5 (14.3)	35 (39.3)	
Systolic pressure (mmHg)	131 (121-145)	132 (122-150)	0.683
≤132	20 (57.1)	46 (51.7)	0.584
>132	15 (42.9)	43 (48.3)	
Diastolic pressure (mmHg)	85 (75-90)	79 (73-87)	0.090
≤80	14 (40.0)	54 (60.7)	0.037
>80	21 (60.0)	35 (39.3)	
Occurrence of ARDS in the course	2 (5.7)	89 (100.0)	<0.001
Treatments			
Nasal/mask oxygen therapy	33 (94.3)	10 (11.2)	<0.001
Mechanical ventilation	2 (5.7)	79 (88.8)	
Hospitalized period (d)	29 (23-35)	15 (9-22)	<0.001
Radiological estimations			

	Discharge group (n=35)	Death events group (n=89)	p-value
Unilateral involvement	1 (2.9)	0 (0)	0.282
Bilateral involvement	34 (97.1)	89 (100)	
DR score	11 (8-15)	14 (7-17)	0.398
≤13	21 (60.0)	44 (49.4)	0.289
>13	14 (40.0)	45 (50.6)	
Laboratory investigations			
WBC ($\times 10^9/L$)	6.55 (4.94-9.43)	8.09 (5.20-11.11)	0.086
≤7.52	22 (62.9)	40 (44.9)	0.073
>7.52	13 (37.1)	49 (55.1)	
Neutrophil ($\times 10^9/L$)	5.09 (3.73-8.26)	7.03 (4.17-10.23)	0.037
≤6.46	25 (71.4)	37 (41.6)	0.003
>6.46	10 (28.6)	52 (58.4)	
Lymphocyte ($\times 10^9/L$)	0.88 (0.66-1.21)	0.56 (0.39-0.83)	0.001
≤0.64	8 (22.9)	55 (61.8)	<0.001
>0.64	27 (77.1)	34 (38.2)	
Hemoglobin (g/L)	126 (120-137)	131 (122-144)	0.066
≤130	22 (62.9)	43 (48.3)	0.144
>130	13 (37.1)	46 (51.7)	
Platelet ($\times 10^9/L$)	197 (143-234)	182 (151-229)	0.407
≤187	15 (42.9)	48 (53.9)	0.267
>187	20 (57.1)	41 (46.1)	
CRP (mg/L)	53.57 (30.28-78.56)	85.86 (51.35-116.83)	0.001
≤77.35	26 (74.3)	37 (41.6)	0.001
>77.35	9 (25.7)	52 (58.4)	
PCT ($\mu g/L$)	0.09 (0.06-0.17)	0.25 (0.12-0.44)	<0.001
≤0.20	26 (74.3)	36 (40.4)	0.001
>0.20	9 (25.7)	53 (59.6)	
TB ($\mu mol/L$)	12.1 (7.9-16.4)	14.1 (9.9-21.3)	0.082
≤13.6	19 (54.3)	43 (48.3)	0.549
>13.6	16 (45.7)	46 (51.7)	
ALT (U/L)	37 (22-78)	37 (25-56)	0.936
≤37	18 (51.4)	45 (50.6)	0.931
>37	17 (48.6)	44 (49.4)	
AST (U/L)	44 (27-73)	47 (36-67)	0.361
≤47	19 (54.3)	45 (50.6)	0.709
>47	16 (45.7)	44 (49.4)	
LDH (U/L)	393 (244-497)	519 (395-634)	0.001
≤481	26 (74.3)	36 (40.4)	0.001
>481	9 (25.7)	53 (59.6)	
Albumin (g/L)	28.8 (26.6-32.0)	28.0 (25.7-30.6)	0.165
≤28.2	15 (42.9)	48 (53.9)	0.267
>28.2	20 (57.1)	41 (46.1)	
Globulin (g/L)	34.7 (30.6-37.8)	33.9 (31.0-38.3)	0.936
≤34.2	17 (48.6)	45 (50.6)	0.842
>34.2	18 (51.4)	44 (49.4)	
Scr ($\mu mol/L$)	70.0 (54.4-88.2)	75.5 (64.3-102.5)	0.147
≤74.1	20 (57.1)	42 (47.2)	0.318
>74.1	15 (42.9)	47 (52.8)	
D-dimer (mg/L)	1.12 (0.43-5.42)	3.97 (0.78-8.00)	0.010
≤3.06	24 (68.6)	39 (43.8)	0.013
>3.06	11 (31.4)	50 (56.2)	
APTT (s)	38.5 (34.4-43.1)	37.4 (32.7-42.3)	0.286
≤38.0	16 (45.7)	46 (51.7)	0.549
>38.0	19 (54.3)	43 (48.3)	
PT (s)	13.7 (12.9-14.4)	14.2 (13.0-15.2)	0.147
≤13.9	21 (60.0)	41 (46.1)	0.163
>13.9	14 (40.0)	48 (53.9)	
Fibrinogen (g/L)	4.68 (3.75-5.19)	4.58 (3.23-5.25)	0.784
≤4.58	17 (48.6)	45 (50.6)	0.842
>4.58	18 (51.4)	44 (49.4)	
cTnI ($\mu g/L$)	9.9 (3.4-57.1)	24.1 (9.8-155.8)	0.006
≤19.3	20 (57.1)	41 (46.1)	0.267
>19.3	15 (42.9)	48 (53.9)	
BNP (ng/L)	48.6 (21.1-119.5)	88.7 (39.6-192.0)	0.029
≤79.3	21 (60.0)	41 (46.1)	0.163
>79.3	14 (40.0)	48 (53.9)	

Abbreviations: CVD: cardiovascular disease; COPD: chronic obstructive pulmonary disease; DR: digital radiograph; WBC: white blood cell count; CRP: C-reactive protein; PCT: procalcitonin; TB: total bilirubin; ALT: alanine aminotransferase; AST: aspartate aminotransferase; LDH: lactate dehydrogenase; Scr: serum creatinine; APTT: activated partial thromboplastin time; PT: prothrombin time; cTnI: cardiac troponin I; BNP: brain natriuretic peptide.

Table 3. Bivariate and multivariate logistic regression of risk factors associated with death events.

	Bivariate		Multivariate	
	OR (95% CI)	p value	OR (95% CI)	p value
Age (y) (>68 vs. ≤68)	1.492 (0.678-3.282)	0.320		
Sex (male vs. female)	2.876 (1.268-6.526)	0.011		
Time of admission after symptom onset (d) (>11 vs. ≤11)	1.281 (0.579-2.835)	0.541		
SpO2 (%) (≤89 vs. >89)	4.253 (1.785-10.133)	0.001	2.959 (1.072-8.167)	0.036
Body temperature (°C) (>38.3 vs. ≤38.3)	1.214 (0.554-2.660)	0.627		
Heart rate (/min) (>89 vs. ≤89)	1.679 (0.759-3.714)	0.201		
Breath rate (/min) (>30 vs. ≤30)	3.889 (1.377-10.979)	0.010		
Systolic pressure (mmHg) (≤132 vs. >132)	1.246 (0.567-2.741)	0.584		
Diastolic pressure (mmHg) (≤80 vs. >80)	2.314 (1.041-5.145)	0.040		
DR score (>13 vs. ≤13)	1.534 (0.694-3.393)	0.291		
WBC (×10 ⁹ /L) (>7.52 vs. ≤7.52)	2.073 (0.929-4.627)	0.075		
Neutrophil (×10 ⁹ /L) (>6.46 vs. ≤6.46)	3.514 (1.508-8.187)	0.004		
Lymphocyte (×10 ⁹ /L) (≤0.64 vs. >0.64)	5.460 (2.226-13.393)	<0.001	4.015 (1.457-11.065)	0.007
Hemoglobin (g/L) (≤130 vs. >130)	0.552 (0.248-1.232)	0.147		
Platelet (×10 ⁹ /L) (≤187 vs. >187)	1.561 (0.709-3.435)	0.268		
CRP (mg/L) (>77.35 vs. ≤77.35)	4.060 (1.706-9.665)	0.002	2.852 (1.054-7.715)	0.039
PCT (µg/L) (>0.20 vs. ≤0.20)	4.253 (1.785-10.133)	0.001	3.554 (1.284-9.834)	0.015
TB (µmol/L) (>13.6 vs. ≤13.6)	1.270 (0.580-2.783)	0.550		
ALT (U/L) (>37 vs. ≤37)	1.035 (0.473-2.264)	0.931		
AST (U/L) (>47 vs. ≤47)	1.161 (0.530-2.544)	0.709		
LDH (U/L) (>481 vs. ≤481)	4.253 (1.785-10.133)	0.001	3.185 (1.154-8.788)	0.025
Albumin (g/L) (≤28.2 vs. >28.2)	1.561 (0.709-3.435)	0.268		
Globulin (g/L) (≤34.2 vs. >34.2)	1.083 (0.495-2.368)	0.842		
Scr (µmol/L) (>74.1 vs. ≤74.1)	1.492 (0.678-3.282)	0.320		
D-dimer (mg/L) (>3.06 vs. ≤3.06)	2.797 (1.223-6.398)	0.015		
APTT (s) (>38.0 vs. ≤38.0)	1.270 (0.580-2.783)	0.550		
PT (s) (>13.9 vs. ≤13.9)	1.756 (0.794-3.886)	0.165		
Fibrinogen (g/L) (≤4.58 vs. >4.58)	1.083 (0.495-2.368)	0.842		
cTnI (µg/L) (>19.3 vs. ≤19.3)	1.561 (0.709-3.435)	0.268		
BNP (ng/L) (>79.3 vs. ≤79.3)	1.756 (0.794-3.886)	0.165		

Abbreviations: DR: digital radiograph; WBC: white blood cell count; CRP: C-reactive protein; PCT: procalcitonin; TB: total bilirubin; ALT: alanine aminotransferase; AST: aspartate aminotransferase; LDH: lactate dehydrogenase; Scr: serum creatinine; APTT: activated partial thromboplastin time; PT: prothrombin time; cTnI: cardiac troponin I; BNP: brain natriuretic peptide.

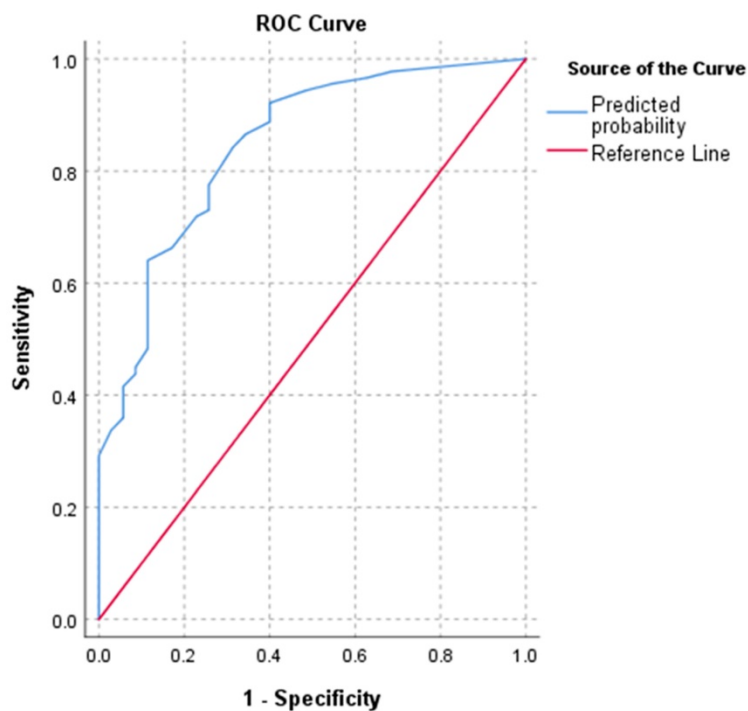


Figure 2. ROC curve of multivariate logistic regression. **Note:** C-index of the predicted probability was 0.845 (p<0.001).

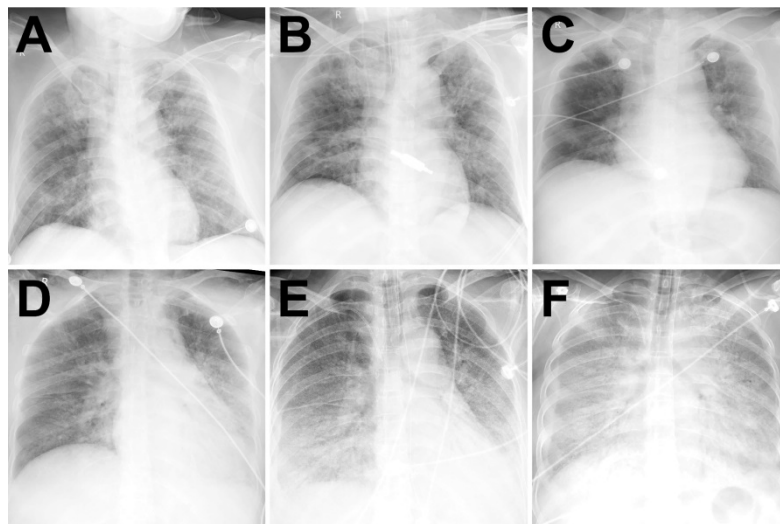


Figure 3. Typical radiographic aggravation manifestations. Note: A-C from a patient in discharge group with fever and cough for six days. After admission, chest DR showed a large area of air-space opacity in the bilateral lungs (DR score: 17 points) (A). Day 6, worsening respiratory failure (hypoxemia and tachypnea) necessitated intubation and ventilation (Critical aggravation). Chest DR demonstrated progressive diffuse opacity of bilateral lungs (DR score: 17 points) (B). Day 11, the intubation was removed and face-mask oxygen treatment was performed. Day 22, chest DR demonstrated partial absorption of the lesions with the reduced density in bilateral lungs (DR score: 14 points) (C). The patient eventually recovered and discharged on Day 38. D-F from a patient in death event group with fever for one week. After admission, chest DR showed a large area of air-space opacity in the left mid-zone and bilaterally lower zones (DR score: 10 points) (D). Day 4, worsening respiratory failure (hypoxemia and tachypnea) necessitated intubation and ventilation (Critical aggravation). Chest DR demonstrated progressive diffuse opacity of bilateral lungs, with bilateral air bronchograms and a right pleural effusion (DR score: 15 points) (E). Day 9, chest DR demonstrated complete opacity of bilateral lungs, with bilateral air bronchograms and a right pleural effusion (DR score: 24 points) (F). The patient eventually died on Day 12 due to refractory respiratory failure.

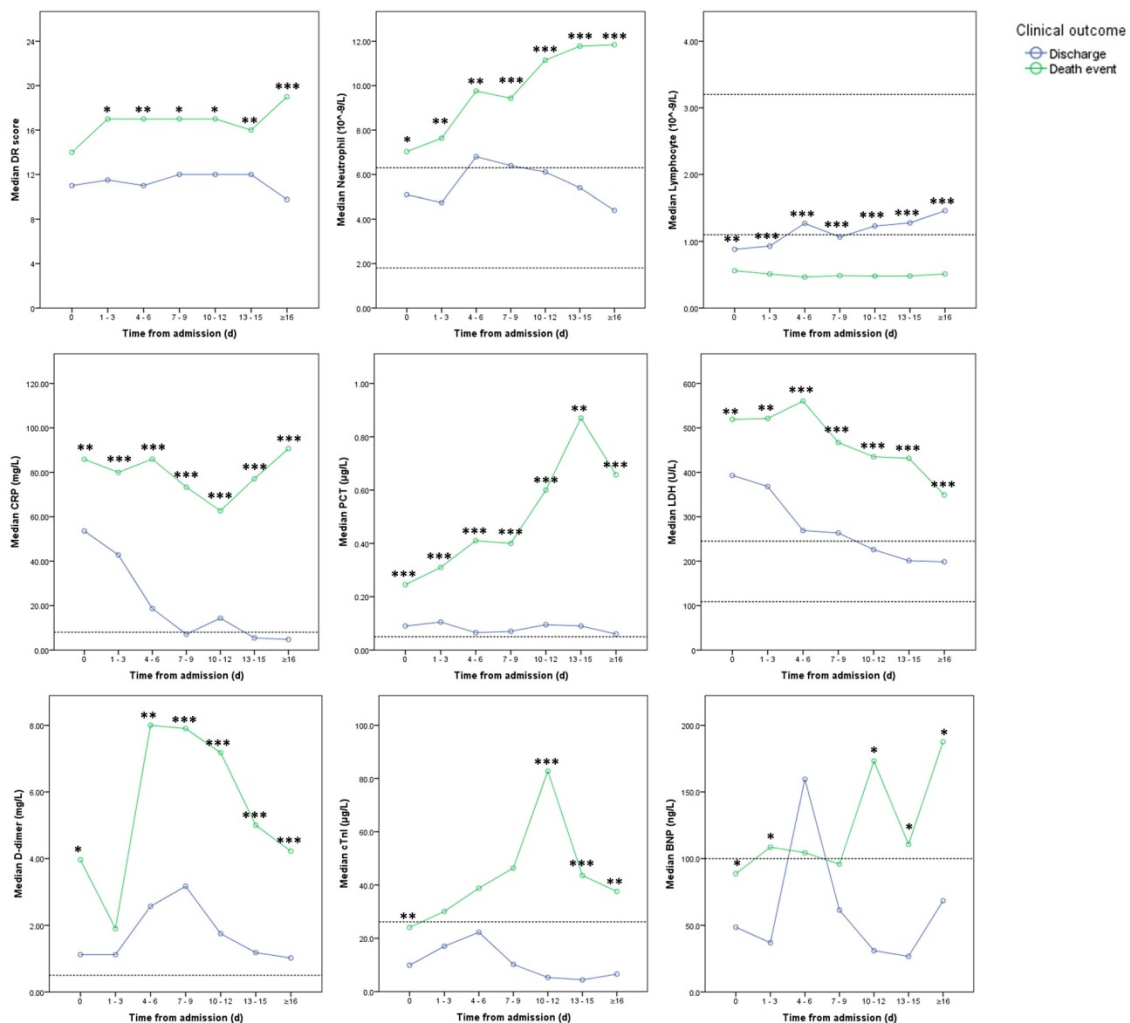


Figure 4. Dynamic changes in DR scores and laboratory parameters between two groups. Note: Imaginary lines indicated the normal reference boundary; Man-Whitney U test (* $p < 0.05$; ** $p < 0.01$; *** $p < 0.001$).

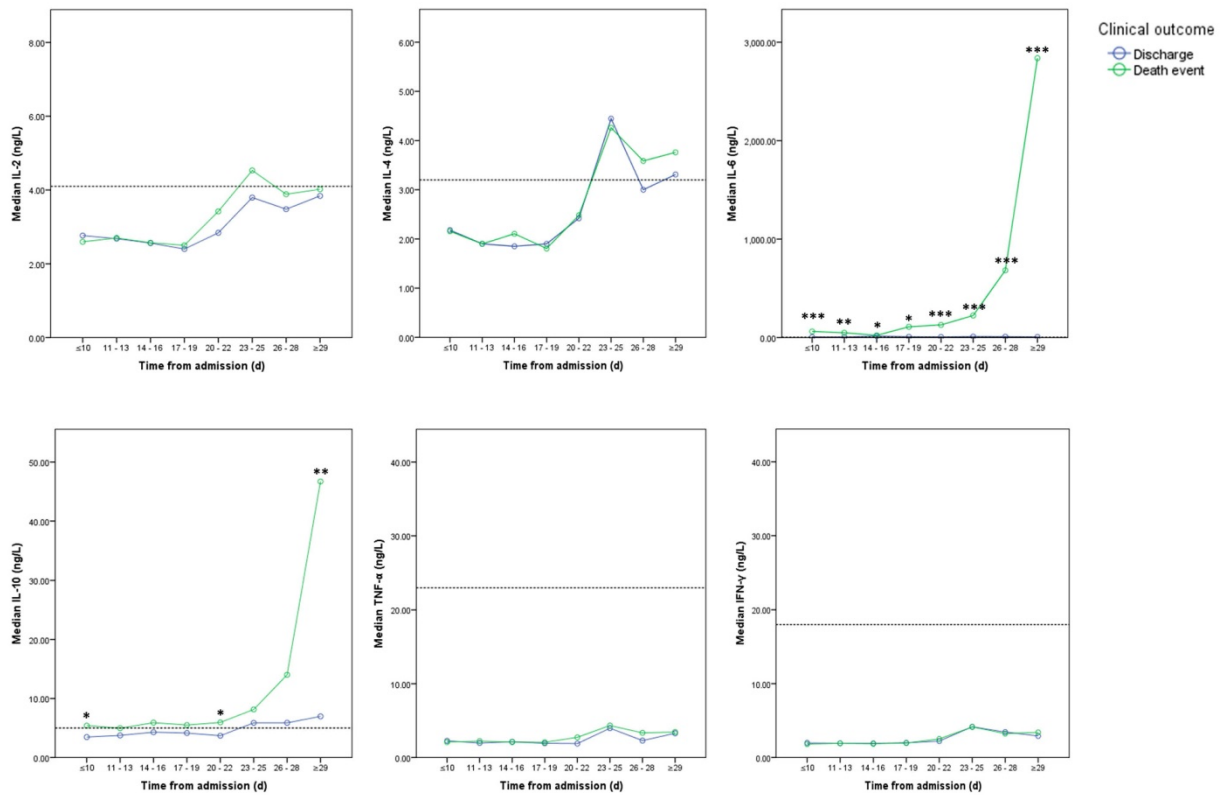


Figure 5. Dynamic changes in cytokine profile between two groups. Note: Imaginary lines indicated the normal reference boundary; Man-Whitney U test (* $p < 0.05$; ** $p < 0.01$; *** $p < 0.001$).

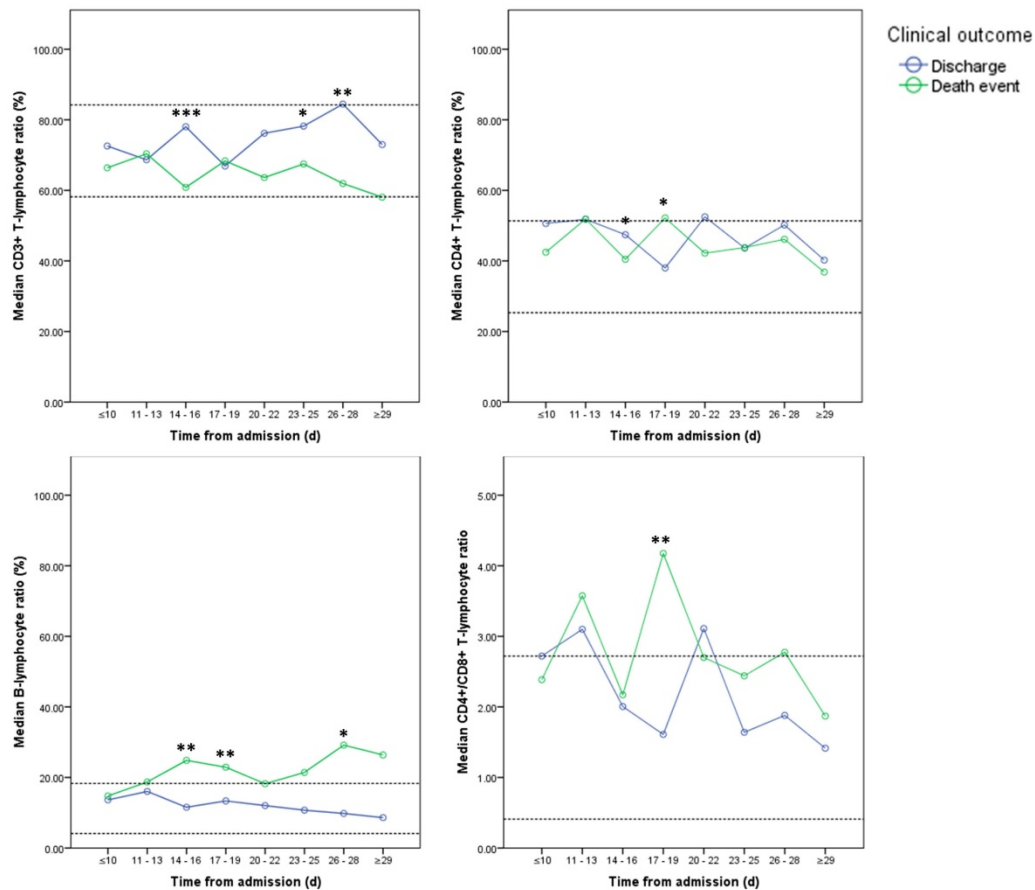


Figure 6. Dynamic changes in Lymphocyte subsets between two groups. Note: Imaginary lines indicated the normal reference boundary; Man-Whitney U test (* $p < 0.05$; ** $p < 0.01$; *** $p < 0.001$).

Up to date, some previous studies already revealed multiple risk factors associated with poor prognosis, such as compromised respiratory status, older age, male sex, lymphocytopenia, high Sequential Organ Failure Assessment score, and elevated levels of CRP, PCT, LDH, and D-dimer [1, 5-8, 10-12]. However, these studies did not consider the impacts of the disease severities on admission in the analysis. As a complementary, this study provided a risk-factor analysis only in the population of severe COVID-19 who presented a higher mortality rate than moderate COVID-19 [8]. As a case-control study, the bivariate logistic regression returned with ten stratified risk factors for death with statistical significances, including male sex, $SpO_2 \leq 89\%$, breath rate $>30/\text{min}$, diastolic pressure $\leq 80\text{mmHg}$, neutrophil $>6.46 \times 10^9/\text{L}$, lymphocyte $\leq 0.64 \times 10^9/\text{L}$, CRP $>77.35\text{mg/L}$, PCT $>0.20\mu\text{g/L}$, LDH $>481\text{U/L}$, and D-dimer $>3.06\text{mg/L}$. Further multivariate logistic regression using the forward-LR method resulted in a significant fitting model with good discriminatory power (C-index=0.845). The results revealed $SpO_2 \leq 89\%$, lymphocyte $\leq 0.64 \times 10^9/\text{L}$, CRP $>77.35\text{mg/L}$, PCT $>0.20\mu\text{g/L}$, and LDH $>481\text{U/L}$ on admission were the independent risk factors, among which the lymphocyte $\leq 0.64 \times 10^9/\text{L}$ demonstrated the highest OR of 4.015. Moreover, the abnormalities of lymphocyte, CRP, PCT, and LDH stayed or continuously aggravated in death event group but gradually recovered to normal levels in discharge group. Therefore, the dynamic monitoring of these laboratory parameters could estimate the prognosis in clinical practice for severe patients.

In further analysis of the lymphocyte subsets in limited groups, no obvious abnormalities of CD3+, CD4+, and CD8+ T-lymphocyte ratio were found in both groups. However, relatively high CD4+/CD8+ T-lymphocyte ratio above or near the upper normal reference was observed in both groups, while it seemed higher in death event group. Therefore, it indicated no obvious impairment of the immune function, though lymphocyte reduced in severe COVID-19. As an evidence, a previous post-mortem autopsy study of COVID-19, it revealed reduced CD4+ and CD8+ T-lymphocyte in peripheral blood with overactivation of T-lymphocyte manifested by an increase of Th17 and high cytotoxicity of CD8+ T-lymphocytes [5]. As a result, severe immune injury could occur in severe patients.

To date, no confirmed evidence of the viral attack on solid organs other than lung was observed in pathological analysis and autopsy [5, 27]. Two previous studies in the same center found significantly elevated levels of multiple serum cytokines in severe COVID-19 patients, such as IL-1 β ,

IL-2, IL-6, TNF- α , and INF- γ , which were associated with the disease severity [3, 7]. It was suspected a phenomenon named "cytokine storm" might result in the progression of the disease in viral pneumonia [28, 29]. In previous studies, COVID-19 was thought to induce "cytokine storm" that then caused multi-organ impairment involving the heart, liver, and kidney [6, 7, 10, 13]. However, the clinical course of our patients did not appear to follow this sequence of events. Among multiple cytokines, only IL-6 as one major proinflammatory factor significantly increased with time in death event group with the simultaneous increase of IL-10 as an anti-inflammatory factor, while IL-2, IL-4, TNF- α , and INF- γ neither showed significant differences between two groups nor demonstrated obvious abnormalities in the course. Besides, the hepatic and renal related parameters were not significantly elevated. Therefore, the result of this study didn't support this fatal hypothesis of "cytokine storm" with induced multi-organ impairments in severe COVID-19. It was speculated that in those published studies, the dynamic changes of the related laboratory parameters were demonstrated as the mean value which might be not appropriate due to the non-normal distribution of the data [6, 10]. Thus, the dynamic demonstration with the median value was more reliable in this study.

On the other hand, CRP, PCT, IL-6, and BNP as the major indicators of inflammatory severity in intensive care medicine, had significantly increased or stayed at a relatively high level in death event group, corresponding to the aggravated sepsis in these patients which could cause life-threatening organ dysfunction [30, 31]. As indirect evidence, increased CD4+/CD8+ T-lymphocyte ratio in death group was also a typical abnormality in sepsis [32]. Meanwhile, gradually increased neutrophil was observed indicating progressive inflammation as part of the inadequate host response to viral aggression [33]. In summary, all of these abnormalities could be ascribed to severe systematic inflammation caused by COVID-19. More than this, the virus-induced inflammation could initiate the activation of the fibrinolytic system and fibrin catabolism resulting in the elevation of the D-dimer level in severe patients which was also observed in this study [6, 11, 34]. As a complication, a persistently high level of LDH, cTnI, and BNP indicated potential cardiac impairment in the course like reported in other studies [7, 13]. However, the elevated levels of D-dimer, cTnI, and BNP fluctuated or decreased in the course but not persistently aggravated, while these parameters progressively deteriorated with time in non-survivors in previous studies [6, 11]. As speculation, these differences might be ascribed to more attention paid

to the cardiac protection and microcirculation improvement for the patients in this cohort.

In brief, it was concluded that the systematic inflammation with induced cardiac dysfunction was the leading cause of death events in severe COVID-19 except for ARDS. Thus, the use of corticosteroids should still be considered for critical patients to prevent severe inflammatory response [15-17]. In one recent study, it was found treatment with methylprednisolone significantly decreased death risk [12]. Serum cytokine profiling and lymphocyte subsets analysis might be useful to guide the immunomodulatory treatment [35].

This case-controlled study has limitations. First, the sample size is limited in this study and not all patients underwent sequential cytokine profile and lymphocyte subsets tests. Second, the study was limited by the heterogeneity of treatments based on the disease status with time, which was impossible to summarize in this cohort. Thus, only the speculation was made to explain the fluctuation of some abnormal laboratory parameters in the course. Third, the dynamic changes of vital signs and oxygen saturation were not presented, because they were continuously monitored in the course and sort of affected by the adjusted treatments. At last, some important subsets of lymphocytes including Th17 and Treg were not measured due to the limitation of the laboratory techniques in this provisional isolation center, thus the identification of the immune function was not comprehensive.

Conclusions

In summary, this study revealed the clinical course of severe COVID-19 patients from admission with different outcomes. Dynamic monitoring of lymphocyte, CRP, PCT, and LDH could be meaningful to predict the prognosis of severe patients. Besides, the fatal feature of COVID-19 is mostly attributed to severe systematic inflammation with induced cardiac dysfunction except for ARDS.

Abbreviations

COVID-19: coronavirus disease-19; SARS: Severe Acute Respiratory Syndrome; MERS: Middle East Respiratory Syndrome; ARDS: acute respiratory distress syndrome; ICU: intensive care unit; DR: digital radiograph; CT: computed tomography; IQR: inter-quartile range; OR: odds ratio; 95%CI: 95% confidence interval; VIF: variance inflation factor; CVD: cardiovascular disease; COPD: chronic obstructive pulmonary disease; DR: digital radiograph; WBC: white blood cell count; CRP: C-reactive protein; PCT: procalcitonin; TB: total bilirubin; ALT: alanine aminotransferase; AST:

aspartate aminotransferase; LDH: lactate dehydrogenase; Scr: serum creatinine; APTT: activated partial thromboplastin time; PT: prothrombin time; cTnI: cardiac troponin I; BNP: brain natriuretic peptide.

Supplementary Material

Supplementary figures and tables.

<http://www.medsci.org/v17p1281s1.pdf>

Acknowledgments

We express our sincere gratitude to the emergency services, nurses, doctors and a lot of medical supports from other provinces for their efforts to combat the COVID-19 outbreak in Wuhan. Besides, we are grateful to Dr. Jiazheng Wang and Dr. Dandan Zheng for many useful discussions through the formation and design of this work.

Funding/Support

This work was supported by the National Natural Science Foundation of China [81873919]; Rapid Response Key Project of COVID-19 in Huazhong University of Science and Technology [2020kfyXGYJ021]; and Key Special Project of Ministry of Science and Technology, China [2020YFC0845700].

Author Contributions

FP, LY, and YL contributed equally to this article. FP, LY, YL, YH, and CZ designed this study and had full access to all data in the study and take responsibility for the integrity of the data and the accuracy of the data analysis. FP, LY, YL, and BL wrote the initial manuscript together with all authors providing critical feedback and edits to subsequent revisions. YL, BL, Lin Li, Lingli Li, TY, DL, and SG carried out the retrieving work and information input including clinical, radiological, and laboratory data from the institutional electronic database. CZ and LY take responsibility for the integrity of the data and the accuracy of the data analysis. CZ and LY were blinded to perform the semi-quantitative analysis of the radiological images on the Vue PACS system. FP and YH finished the statistical data analysis, and LY, BL, LY, and CZ performed the proofreading work. All authors approved the final draft of the manuscript and agreed to be accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved.

Competing Interests

The authors have declared that no competing interest exists.

References

1. The Novel Coronavirus Pneumonia Emergency Response Epidemiology Team. The epidemiological characteristics of an outbreak of 2019 novel coronavirus diseases (COVID-19) in China. *Chinese Journal of Epidemiology*. 2020; 41: 145-51.
2. World Health Organization. Novel coronavirus (2019-nCoV). Situation report 44. Geneva, Switzerland: World Health Organization; 2020. . Published on March 4, 2020; https://www.who.int/docs/default-source/coronaviruse/situation-reports/20200304-sitrep-44-covid-19.pdf?sfvrsn=783b4c9d_2.
3. Chen N, Zhou M, Dong X, Qu J, Gong F, Han Y, et al. Epidemiological and clinical characteristics of 99 cases of 2019 novel coronavirus pneumonia in Wuhan, China: a descriptive study. *Lancet*. 2020; 395: 507-13.
4. World Health Organization. Novel coronavirus (2019-nCoV). Situation report 77. Geneva, Switzerland: World Health Organization; 2020. . Published on April 6, 2020; https://www.who.int/docs/default-source/coronaviruse/situation-reports/20200406-sitrep-77-covid-19.pdf?sfvrsn=21d1e632_2.
5. Xu Z, Shi L, Wang Y, Zhang J, Huang L, Zhang C, et al. Pathological findings of COVID-19 associated with acute respiratory distress syndrome. *The Lancet Respiratory Medicine*. 2020; (Online first).
6. Wang D, Hu B, Hu C, Zhu F, Liu X, Zhang J, et al. Clinical Characteristics of 138 Hospitalized Patients With 2019 Novel Coronavirus-Infected Pneumonia in Wuhan, China. *JAMA*. 2020.
7. Huang C, Wang Y, Li X, Ren L, Zhao J, Hu Y, et al. Clinical features of patients infected with 2019 novel coronavirus in Wuhan, China. *Lancet*. 2020.
8. Guan W-J, Ni Z-Y, Hu Y, Liang W-H, Ou C-Q, He J-X, et al. Clinical Characteristics of Coronavirus Disease 2019 in China. *The New England journal of medicine*. 2020; 10.1056/NEJMoa2002032.
9. Pan F, Ye T, Sun P, Gui S, Liang B, Li L, et al. Time Course of Lung Changes On Chest CT During Recovery From 2019 Novel Coronavirus (COVID-19) Pneumonia. *Radiology*. 2020; 200370.
10. Yang X, Yu Y, Xu J, Shu H, Xia J, Liu H, et al. Clinical course and outcomes of critically ill patients with SARS-CoV-2 pneumonia in Wuhan, China: a single-centered, retrospective, observational study. *The Lancet Respiratory Medicine*. 2020.
11. Zhou F, Yu T, Du R, Fan G, Liu Y, Liu Z, et al. Clinical course and risk factors for mortality of adult inpatients with COVID-19 in Wuhan, China: a retrospective cohort study. *The Lancet*. 2020.
12. Wu C, Chen X, Cai Y, Xia J, Zhou X, Xu S, et al. Risk Factors Associated With Acute Respiratory Distress Syndrome and Death in Patients With Coronavirus Disease 2019 Pneumonia in Wuhan, China. *JAMA Intern Med*. 2020.
13. Ruan Q, Yang K, Wang W, Jiang L, Song J. Clinical predictors of mortality due to COVID-19 based on an analysis of data of 150 patients from Wuhan, China. *Intensive Care Med*. 2020.
14. Bauchner H, Golub RM, Zylke J. Editorial Concern-Possible Reporting of the Same Patients With COVID-19 in Different Reports. *Jama*. 2020.
15. National Health Commission of the People's Republic of China. Diagnosis and treatment protocols of pneumonia caused by a novel coronavirus (trial version 6). Published on February 18, 2020; <http://www.nhc.gov.cn/yzygj/s7653p/202002/8334a8326dd94d329df351d1da8aefc2/files/b218cf6b1bc54639af227f922bf6b817.pdf>.
16. National Health Commission of the People's Republic of China. Diagnosis and treatment protocols of pneumonia caused by a novel coronavirus (trial version 7). Published on March 3, 2020; <http://www.nhc.gov.cn/yzygj/s7653p/202003/46c9294a7dfe4cef80dc7f5912eb1989/files/ce3e6945832a438ea4e415350a8ce964.pdf>.
17. World Health Organization. Clinical management of severe acute respiratory infection when novel coronavirus (nCoV) infection is suspected. Published on March 13, 2020; <https://apps.who.int/iris/rest/bitstreams/1272156/retrieve>.
18. World Health Organization. IMAI district clinician manual: hospital care adolescents and adults : guidelines for the management of illnesses with limited-resources. 2012; https://apps.who.int/iris/bitstream/handle/10665/77751/9789241548290_Vol2_eng.pdf?sequence=3: 111-5.
19. Antonio GE, Ooi CG, Wong KT, Tsui EL, Wong JS, Sy AN, et al. Radiographic-clinical correlation in severe acute respiratory syndrome: study of 1373 patients in Hong Kong. *Radiology*. 2005; 237: 1081-90.
20. Cha MJ, Chung MJ, Kim K, Lee KS, Kim TJ, Kim TS. Clinical implication of radiographic scores in acute Middle East respiratory syndrome coronavirus pneumonia: Report from a single tertiary-referral center of South Korea. *European journal of radiology*. 2018; 107: 196-202.
21. Force ADT, Ranieri VM, Rubenfeld GD, Thompson BT, Ferguson ND, Caldwell E, et al. Acute respiratory distress syndrome: the Berlin Definition. *JAMA*. 2012; 307: 2526-33.
22. World Health Organization. Clinical management of severe acute respiratory infection when novel coronavirus (nCoV) infection is suspected: interim guidance. Published on January 12, 2020; https://www.who.int/docs/default-source/coronaviruse/clinical-management-of-novel-cov.pdf?sfvrsn=bc7da517_2.
23. Akinwande MO, Dikko HG, Samson A. Variance Inflation Factor: As a Condition for the Inclusion of Suppressor Variable(s) in Regression Analysis. *Open Journal of Statistics*. 2015; Vol.05No.07: 14.
24. Jacobi A, Chung M, Bernheim A, Eber C. Portable chest X-ray in coronavirus disease-19 (COVID-19): A pictorial review. *Clin Imaging*. 2020; 64: 35-42.
25. Reissig A, Copetti R, Mathis G, Mempel C, Schuler A, Zechner P, et al. Lung Ultrasound in the Diagnosis and Follow-up of Community-Acquired Pneumonia: A Prospective, Multicenter, Diagnostic Accuracy Study. *Chest*. 2012; 142: 965-72.
26. Chavez MA, Shams N, Ellington LE, Naithani N, Gilman RH, Steinhoff MC, et al. Lung ultrasound for the diagnosis of pneumonia in adults: a systematic review and meta-analysis. *Respiratory Research*. 2014; 15: 50-.
27. Liu X, Wang RS, Qu GQ, Wang YY, Liu P, Zhu YZ, et al. A gross observational report of systematic autopsy in COVID-19 pneumonia. *Journal of Forensic Medicine*. 2020; 36: 19-21.
28. Rommel MGE, Milde C, Eberle R, Schulze H, Modlich U. Endothelial-platelet interactions in influenza-induced pneumonia: A potential therapeutic target. *Anat Histol Embryol*. 2019.
29. Wong CK, Lam CWK, Wu AKL, Ip WK, Lee NLS, Chan IHS, et al. Plasma inflammatory cytokines and chemokines in severe acute respiratory syndrome. *Clin Exp Immunol*. 2004; 136: 95-103.
30. Yang J, Yang Y, Liu Q. [Study on the levels of main inflammatory factors in septic shock and their clinical significance]. *Zhonghua wei zhong bing ji jiu yixue*. 2019; 31: 680-3.
31. Singer M, Deutschman CS, Seymour CW, Shankar-Hari M, Annane D, Bauer M, et al. The Third International Consensus Definitions for Sepsis and Septic Shock (Sepsis-3). *Jama*. 2016; 315: 801-10.
32. Tschaikowsky K, Hedwig-Geissing M, Schiele A, Bremer F, Schywalsky M, Schüttler J. Coincidence of pro- and anti-inflammatory responses in the early phase of severe sepsis: Longitudinal study of mononuclear histocompatibility leukocyte antigen-DR expression, procalcitonin, C-reactive protein, and changes in T-cell subsets in septic and postoperative patients. *Critical care medicine*. 2002; 30: 1015-23.
33. Stiel L, Meziani F, Helms J. Neutrophil Activation During Septic Shock. *Shock (Augusta, Ga)*. 2018; 49: 371-84.
34. Arslan S, Ugurlu S, Bulut G, Akkurt I. The association between plasma D-dimer levels and community-acquired pneumonia. *Clinics (Sao Paulo)*. 2010; 65: 593-7.
35. Chen L, Liu HG, Liu W, Liu J, Liu K, Shang J, et al. [Analysis of clinical features of 29 patients with 2019 novel coronavirus pneumonia]. *Zhonghua Jie He He Hu Xi Za Zhi*. 2020; 43: E005.