

2020; 17(18): 3039-3048. doi: 10.7150/ijms.47807

Research Paper

Vitamin D Status and Pregnancy Complications: Serum 1,25-di-hydroxyl-Vitamin D and its Ratio to 25-hydroxy-Vitamin D are Superior Biomarkers than 25-hydroxy-Vitamin D

Ibrahim A. Albahlol^{1,2}, Abdulrahman H. Almaeen³, Abdulrahman A. Alduraywish⁴, Umar F. Dar⁵, Tarek H. El-Metwally^{6,7⊠}

- 1. Department of Obstetrics and Gynecology, College of Medicine, Jouf University, Sakaka, Saudi Arabia. iaelbahloul@ju.edu.sa
- 2. Department of Obstetrics and Gynecology, Mansoura University, Mansoura, Egypt.
- 3. Department of Pathology, College of Medicine, Jouf University, Sakaka, Saudi Arabia. ahalmaeen@ju.edu.sa
- 4. Department of Internal Medicine, College of Medicine, Jouf University, Sakaka, Saudi Arabia. dr-aaad@ju.edu.sa
- 5. Department of Family and Community Medicine, College of Medicine, Jouf University, Sakaka, Saudi Arabia. ufdar@ju.edu.sa
- Department of Pathology, Biochemistry Division, College of Medicine, Jouf University, Sakaka, Saudi Arabia. thelmetwally@ju.edu.sa.
- 7. Department of Medical Biochemistry, Faculty of Medicine, Assiut University, Assiut, Egypt.

Corresponding author: Prof. Dr. Tarek H. El-Metwally, PhD, Department of Pathology, Biochemistry Division, College of Medicine, Jouf University, 2004 Sakaka 42421, Saudi Arabia. 00966-541860565. thelmetwally@ju.edu.sa. http://orcid.org/0000-0001-9040-6642

© The author(s). This is an open access article distributed under the terms of the Creative Commons Attribution License (https://creativecommons.org/licenses/by/4.0/). See http://ivyspring.com/terms for full terms and conditions.

Received: 2020.05.05; Accepted: 2020.09.14; Published: 2020.10.18

Abstract

Vitamin D (VitD) deficiency during pregnancy has been associated with adverse neonatal outcomes and increased risk of late pregnancy complications. We planned to correlate serum VitD biomarkers; 25-hydroxyvitamin D (25-OH-VitD) and 1,25-dihydroxyvitamin D (1,25-diOH-VitD) levels; and their ratio with the frequency of feto-maternal pregnancy complications. A prospective cross-sectional case-control study was conducted at Aljouf Maternity and Children Hospital, Sakaka, Saudi Arabia, during the period of September 1, 2017 to September 30, 2019. 322 pregnant women were stratified into 2 groups: controls (110 cases) and complicated group (212 cases). The later comprised severe preeclamptic toxemia associated with intrauterine growth restriction (58 cases), gestational diabetes mellitus (GDM; 82 cases), abortion (26 cases), undisturbed ectopic pregnancy (16 cases), premature rupture of membranes (PROM; 14 cases), and, inevitable preterm labour (16 cases). After clinical assessment, peripheral blood samples were collected. Serum biomarkers were measured using specific immunoassays. The direct 1,25-diOH-VitD/25-OH-VitD ratio was calculated. Serum 25-OH-VitD indicated widely spreading VitD deficiency among participants with significantly higher levels in controls vs. GDM subgroup only. 1,25-diOH-VitD levels and the ratio were markedly reduced in the six complicated subgroups vs. controls, with non-significant differences amongst the complicated subgroups. ROC analysis showed very high sensitivity and specificity, to differentiate patients from controls, only for 1,25-diOH-VitD (AUC = 0.965; 0.947 - 0.983, p < 0.001) followed by the ratio but not 25-OH-VitD. In conclusions, 25-OH-VitD did not show significant changes except for GDM. 1,25-diOH-VitD levels and the ratio showed strong associations with pregnancy complications. Serum 1,25-di-OH-VitD and its ratio to 25-OH-VitD are more reliable and physiologically relevant biomarkers for VitD status in pregnancy.

Key words: Serum vitamin D biomarkers, pregnancy outcomes, pregnancy complications, 25-hydroxyvitamin D, 1,25-dihydroxyvitamin D

Introduction

Vitamin D (VitD) deficiency became a global epidemic, particularly for women of reproductive age.

Nuclear receptor-mediated, the dihydoxy active form of VitD, 1,25-diOH-VitD, controls cellular

proliferation, differentiation and apoptosis through targeting ≥1000 genes. Nevertheless, pathogenetic role of hypovitaminosis D in a number of comorbidities is still hotly debated [1,2]. With increased requirements of the conditional VitD, pregnancy is a high-risk factor for VitD deficiency since the fetus gains its VitD requirements from mother [3]. Despite abundant sunlight, published reports confirmed a wide spread of hypovitaminosis D among Saudi citizens [4]. More than half of UK population is VitD deficient, i.e., having serum 25-hydroxy-VitD (25-OH-VitD) <20 ng/mL - as defined by the Endocrine Society [5]. To maintain healthy musculoskeletal system, the Advisory Committee on Scientific Nutrition Department recommended a serum 25-OH-VitD cut-off value of 10 ng/mL for all persons [6].

Linked to hypovitaminosis D, adverse fetal outcomes included abortion, intrauterine growth restriction, fetal death and congenital malformations. Maternal adverse effects involved preeclampsia, gestational diabetes mellitus (GDM) and increased risk of preterm labor [7-9]. Although the causative role of VitD deficiency in different pregnancy complications is still an open question - due to the insufficient reports locally and globally, VitD supplementation may reduce GDM, preeclampsia, preterm labor, and subnormal neonatal anthropometric measures [10]. Meta-analyses of observational studies correlate hypovitaminosis D with pregnancy complications without asserting causation [11]. The longitudinal pattern of changes in 25-OH-VitD concentrations and its relationship with 1,25-diOH-VitD throughout pregnancy remains largely unclear. Time-dependent decreases, increases or no change in 25-OH-VitD levels were reported, while 1,25-diOH-VitD levels mostly increase at term compared to non-pregnant women. Additionally, serum VitD cutoffs that are defined for the general adult population are inappropriately used for pregnancy [3].

We planned to assess the relationship between hypovitaminosis D and feto-maternal pregnancy complications and morbidities, using the more patho-physiologically relevant biomarkers. We measured serum 25-OH-VitD, as the established VitD biomarker, and, 1,25-diOH-VitD as a functional VitD biomarker, and their ratio, and, correlated them with the existing pregnancy feto-maternal status.

Materials and Methods

Settings and participants

This prospective cross-sectional case control study was carried out at Aljouf Maternity and Children Hospital, Sakaka, Saudi Arabia, during the period between September 1, 2017 and September 30, 2019. It was approved by the local bioethical committees of Jouf University and Ministry of Health (#6-16-4/40). Written informed consents were obtained. The exclusion criteria were uncertain diagnosis, obesity, anemia, endocrinal disorders (hypo-/hyper-parathyroidism and thyroidism, and diabetes other than GDM), immobilization for any reason, on anti-convulsion drug, acute infections, malabsorption syndromes, comorbid chronic medical disorder (renal or liver impairment, inflammatory and immunological disorders), multiple pregnancy, hemorrhage superimposed antepartum and preeclampsia.

Examination and investigations

Among 450 pregnant women attending the hospital, 322 voluntary participants fulfilling the inclusion criteria were recruited. The 322 participants were classified into 2 groups; control group of 110 women who had normal feto-maternal pregnancy, and, complicated pregnancy group comprising 212 participants. The latter group was further subdivided into: severe preeclamptic toxemia (PET) associated with intrauterine growth restriction (IUGR; 58 cases), GDM (82 cases), abortion (26 cases), undisturbed ectopic pregnancy (16 cases), premature rupture of membranes (PROMs; 14 cases), and, inevitable preterm labor (16 cases).

After history taking (for age, gravidity, parity, duration of pregnancy in weeks and significant medical history) and general examination (for vital signs and weight and height to calculate body mass index; BMI), the head, face, neck, chest, heart, back, lower limbs and abdomen were systematically examined. The pregnancy was assessed ultra-sonographically for viability and fetal biometry to determine pregnancy duration and the expected fetal weight, condition of amniotic fluid and for diagnosing fetal anomalies, if present. Women with routine laboratory findings that contradict the inclusion criteria were excluded.

Serum was recovered from peripheral blood samples by centrifugation and was aliquotted and stored till used at -80 °C. Specific quantitative ELISA immunoassay kits (Sunlong Biotech Co. Ltd., Zhejiang, China) were used to measure total 25-OH-VitD (in ng/mL; cat# SL2762Hu) and 1,25-DiOH-VitD (in pg/mL; cat# SL2845Hu). The 1,25-DiOH-VitD/25-OH-VitD direct ratio was calculated for each patient. Participants were stratified using the clinically relevant 25-OH-VitD cutoff levels of high/toxic as $\geq 50/>80$ ng/mL, normal as \geq 30-50 ng/mL, insufficient as \geq 20 - <30 ng/mL, deficient as ≥10 - <20 ng/mL and severely deficient as <10 ng/mL [5, 12-14]. Unfortunately, all of the cutoff recommendations do not refer to pregnancy and its complication as distinct clinical entities.

Statistical analysis

Data was analyzed using SPSS (Statistical Package for Social Sciences, Version 23.0. Armonk, NY: IBM Corp). We expressed descriptive qualitative data as frequency and percentage, and, quantitative data as range and mean \pm standard deviation (SD). Normal distribution was checked bv Kolmogorov-Sminov test. One-way ANOVA with Tukey's post-test was employed for multiple comparisons. Correlation among variables was assessed by Spearman correlation coefficient test. Receiver Operating Characteristic (ROC) curve analysis was used to determine the area under curve (AUC) for 25-OH-VitD, 1,25-DiOH-VitD and their ratio to check their sensitivity and specificity to differentiate the cases from controls. P value of <0.05 at a confidence level of 95% was considered significant.

Results

Demographics, anthropometrics, clinical characteristics and vitamin D biomarkers (Table 1)

Age was nonsignificantly different among all groups except for preeclamptics vs. each of controls (p = 0.011) and PROMs (p = 0.014), and, PROMs vs. GDM (p = 0.035). BMI showed nonsignificant difference among the investigated groups except comparing GDM vs. each of controls (p = 0.002), abortion (p = 0.02), ectopic pregnancy (p < 0.001) and

PROMs (p <0.001); and preeclamptics vs. each of ectopic pregnancy (p < 0.028) and PROM (p = 0.045). Gravidity showed nonsignificant difference among the investigated groups except comparing GDM vs. each of controls (p < 0.001), preeclamptics (p < 0.001), abortion (p <0.036), ectopic pregnancy (p = 0.002) and PROMs (p = 0.002). Parity showed nonsignificant difference among the investigated groups except comparing GDM vs. each of preeclamptics (p < 0.001), abortion (p = 0.035), ectopic pregnancy (p = 0.002) and PROMs (p <0.035). Duration of pregnancy showed very strong significant difference among the investigated groups with a few non-significant exceptions; it was significantly different comparing controls vs. all complication subgroups (p < 0.01 -0.001) except GDM, comparing preeclamptics vs. all other subgroups (p <0.05 - <0.001) except preterm labour, comparing GDM vs. the others (p <0.001), comparing abortion vs. the others (p < 0.001) except ectopic pregnancy, and, comparing ectopic pregnancy vs. the others (p < 0.001) except PROMs.

Serum 25-OH-VitD levels showed nonsignificant difference among the investigated groups except comparing GDM vs. each of controls (p < 0.001) and ectopic pregnancy (p = 0.01). Serum 1,25-diOH-VitD levels showed nonsignificant difference among the investigated groups of patients. However, controls had markedly higher levels vs. each of the six complicated groups (p < 0.001). Similarly, the direct 1,25-diOH-VitD/25-OH-VitD ratio showed very marked significant reduction comparing controls vs. each of the six patients' groups (p < 0.001), albeit, with nonsignificant difference among the complicated groups.

Table 1. Demographic, anthropometric, clinical characteristics, serum 25-hydoxyvitamin D (25-OH-VitD) and 1,25-dihydroxyvitamin D (1,25-DiOH-VitD), and their ratio in Saudi women with and without pregnancy complications.

Parameter	Controls	Preeclampsia	GDM	Abortion	EP	PROMs	PTL
	n = 110	n = 58	n = 82	n = 26	n = 16	n = 14	n = 16
Age, Years	28.51 ± 5.538	31.52 ± 5.202	31.6 ± 5.7	31.2 ± 4.76	27.1 ± 6.27	26.1 ± 4.82	28.5 ± 4.41
	(18 - 40)	(19 - 42)	(22 - 49)	(23 - 42)	(21 - 40)	(19 - 32)	(20 - 34)
BMI, kg/m ²	22.21 ± 1.786	22.6 ± 1.706	23.3 ± 1.93	21.9 ± 2.06	20.9 ± 2.35	20.9 ± 2.35	22.5 ± 2.02
	(17.8 - 25.1)	(18.8 - 25.2)	(18.2 - 25)	(17.8 - 25.1)	(17.6 - 25.4)	(18.4 - 24.2)	(18.6 - 24.6)
Gravidity, n	4.055 ± 2.365	3.345 ± 1.617	5.39 ± 2.07	3.92 ± 1.81	3.13 ± 1.67	3 ± 2.72	4 ± 2.42
-	(1 - 10)	(1 - 6)	(1 - 12)	(1 - 8)	(1 - 6)	(1 - 7)	(1 - 8)
Parity, n	2.691 ± 2.208	2.034 ± 1.578	3.37 ± 1.55	2.08 ± 1.57	1.38 ± 1.45	1.71 ± 2.27	2.13 ± 1.96
-	(0 - 8)	(0 - 5)	(0 - 8)	(0 - 5)	(0 - 4)	(0 - 5)	(0 - 6)
Pregnancy Duration, Weeks	35.45 ± 5.465	31.17 ± 4.849	25.1 ± 9.68	11.7 ± 4.09	6 ± 0.73	37.1 ± 2.74	33.6 ± 1.93
	(20 - 41)	(22 - 37)	(6 - 40)	(7 - 18)	(5 - 7)	(33 - 39)	(29 - 35)
25-OH-VitD, ng/mL	28.46 ± 22.77	23.02 ± 3.674	17.7 ± 4.6	23.9 ± 19.5	33.6 ± 37	20 ± 2.59	20.8 ± 1.17
-	(13.48 - 105.6)	(15.17 - 35.97)	(11 - 35.5)	(15.7 - 90.1)	(17.5 - 128)	(17 - 24.7)	(18.5 - 22.6)
1,25-DiOH-VitD, pg/mL	111.3 ± 47.54	45.04 ± 9.178	42.4 ± 12.0	45.0 ± 22.4	35.8 ± 7.29	56.6 ± 52.3	34.0 ± 4.29
	(49.2 - 281.3)	(23.3 - 63.2)	(15.9 - 84.0)	(32.0 - 119.7)	(27.3 - 50.8)	(25.8 - 179.3)	(27.8 - 41.1)
1,25-DiOH-VitD/25: OH-VitD Ratio	4.956 ± 2.650	1.998 ± 0.533	2.53 ± 0.994	2.34 ± 1.41	1.65 ± 0.72	2.73 ± 2.24	1.63 ± 0.2
	(0.956 - 12.9)	(1.215 - 3.582)	(0.94 - 6.24)	(0.485 - 6.71)	(0.235 - 2.9)	(1.52 - 7.96)	(1.34 - 2.0)

Data shown are number of participants per group (n) and mean ± SDM and range. GDM = Gestational diabetes, EP= Ectopic pregnancy, PROMs = Premature rupture of membranes, PTL = Preterm labor. For significance of differences, see the text.

Stratification of our participants on bases of the clinical 25-OH-VitD cutoff values (Table 2)

Using serum 25-OH-VitD as the biomarker for VitD status, 43.636% of controls were deficient (≥ 10 ng/mL), 38.181% were insufficient (≥ 20 ng/mL), and, equal proportions (9.09%) were either normal (≥30 ng/mL) or having toxic levels (>80 ng/mL). 82.759% of preeclamptic cases were insufficient, 13.793% were deficient and 3.448% had normal levels. 82.927% of GDM cases were deficient, 12.195% were insufficient and 4.878 had normal levels. 84.615% of abortion cases deficient, and equal proportions (7.692%) had either insufficient or toxic levels. 50.0% of ectopic pregnancy cases were deficient, 37.5% were insufficient and the rest (12.5%/2) had toxic levels. 71.429%/ of PROMs cases were deficient and 28.571% were insufficient. 75% of preterm labor cases were insufficient and 25.0% were deficient. Fortunately, severe deficiency (<10 ng/mL) was not observed in our cases. Therefore, VitD insufficiency/deficiency is widely spreading among the study participants (292 out of 322, i.e., 90.683%). Controls presented a better picture only considering insufficiency/deficiency proportion, and, % of normal/super-normal levels.

Table 2. Serum 25-hydroxyvitamin D levels among Saudi women with and without pregnancy complications.

Group	Toxic (>80)	Normal (≥30 - <50)	Insufficiency (≥20 - <30)	Deficiency (≥10 - <20)	Insf./Defi.
Controls (n = 110)	10 (9.09)	10 (9.09)	42 (38.181)	48 (43.636)	90 (81.818)
Preeclamptics (n = 58)	0 (0)	2 (3.448)	48 (82.759)	8 (13.793)	56 (96.552)
Gestational diabetes (n = 82)	0 (0)	4 (4.878)	10 (12.195)	68 (82.927)	78 (95.122)
Abortion (n = 26)	2 (7.692)	0 (0)	2 (7.692)	22 (84.615)	24 (92.308)

Group	Toxic (>80)	Normal (≥30 - <50)	Insufficiency (≥20 - <30)	Deficiency (≥10 - <20)	Insf./Defi.
Ectopic Pregnancy (n = 16)	2 (12.5)	0 (0)	6 (37.5)	8 (50.0)	14 (87.5)
PROMs (n = 14)	0 (0)	0 (0)	4 (28.571)	10 (71.429)	14 (100)
Preterm labor (n = 16)	0 (0)	0 (0)	12 (75.0)	4 (25.0)	16 (100)
Total (n = 322)	14 (4.508)	16 (4.969)	124 (38.509)	168 (52.174)	292 (90.683)

Data shown are frequency; n and (%). Insf. = Insufficiency, Defi. = Deficiency. For significance of differences, see the text.

ROC curve analysis

ROC curve analysis showed that 1,25-DiOH-VitD level is the most sensitive and most specific biomarker for differentiating between cases and controls; with an AUC of 0.965 (0.947 - 0.983, p <0.001). It is followed by 1,25-DiOH-VitD25-OH-VitD ratio; with AUC of 0.843 (0.791 - 0.676, p <0.001). 25-OH-VitD levels fell near the diagonal line with no effect, i.e., showing least sensitivity and least specificity (AUC = 0.61; 0.543 - 0.676, and, p = 0.001) (Figure 1 and Table 3).

Table 3. Area Under the Curve (AUC) for differentiation between cases (n = 212) and controls (n = 110) using each of 1,25-diOH-VitD, 25-OH-VitD: 1,25-diOH-VitD ratio, and, of 25-OH-VitD among Saudi women with and without pregnancy complications.

Test Result Variable(s)	AUC P value	e* Range of AUC	Range of AUC		
		Lower Bound	Upper Bound		
1,25-diOH-VitD	0.965 <0.001	0.947	0.983		
1,25-diOH-VitD: 25-OH-VitD ratio	0.843 < 0.001	0.791	0.895		
25-OH-VitD	0.610 0.001	0.543	0.676		

*Null hypothesis: true area = 0.5. Data shown are AUC, p values and AUC range at 95% Confidence Interval.



Figure 1. The ROC curve for sensitivity and specificity of each of 1,25-DiOH-VitD levels, 1,25-DiOH-VitD: 25-OH-VitD ratio, and, 25-OH-VitD levels for differentiation between cases (n = 212) and controls (n = 110) among Saudi women with and without pregnancy complications. Diagonal segments were produced by ties.

Results of the correlation analysis (presented as r/p values)

For controls, age showed positive correlations each of gravidity (0.514/<0.001), parity VS. (0.468/<0.001) and BMI (0.837/<0.001), and negative relationship with the ratio (-0.155/<0.05). Gravidity showed positive relationship vs. parity (0.958/<0.001)and BMI (0.480/<0.001). Parity correlated positively with BMI (0.448/<0.001). BMI correlated negatively vs. the ratio (-0.184/= 0.027). Pregnancy duration correlated negatively with 1,25-DiOH-VitD (-0.241/<0.006)(-0.192/=and ratio 0.022). 25-OH-VitD correlated positively vs. 1,25-DiOH-VitD (0.157 < 0.05)and negatively vs. the ratio (-0.647/<0.001). 1,25-DiOH-VitD correlated positively with the ratio (0.562 / < 0.001).

For preeclamptic cases, age had positive correlation vs. gravidity (0.682/<0.001), parity (0.608/<0.001) and BMI (0.690/<0.001), and, negative correlation vs. 1,25-DiOH-VitD (-0.267/= 0.021). Gravidity related had positively with parity (0.497/<0.001), (0.962/<0.001)and BMI but, negatively vs. 1,25-DiOG-VitD (-0.282/= 0.016). Parity correlated positively vs. BMI (0.424/<0.001), and, negatively with 1,25-DiOH-VitD (-0.218/<0.05). BMI correlated positively vs. duration of pregnancy (0.333/= 0.005) and negatively vs. 1,25-DiOH-VitD (-0.275/= 0.017). 25-OH-VitD correlated negatively with the ratio (-0.490/<0.001). 1,25-DiOH-VitD correlated positively vs. the ratio (0.853/<0.001).

For GDM cases, age correlated positively vs. each of gravidity (0.680/<0.001),parity (0.732/<0.001), BMI (0.657/<0.001) and 25-OH-VitD (0.239/= 0.015), but, negatively vs. each of 1,25-DiOH-VitD (-0.179/<0.05) and the ratio (-0.320/<0.002). Gravidity correlated positively with parity (0.865/<0.001), BMI (0.671/<0.001) and 25-OH-VitD (0.216/<0.026), and, negatively vs. duration of pregnancy (-0.294/<0.004) and the ratio (-0.183/<0.05). Parity correlated positively vs. BMI (0.725/<0.001) and 25-OH-VitD (0.187/0.047), and, negatively vs. duration of pregnancy (-0.179/<0.05). positively vs. 25-OH-VitD BMI correlated (0.412/<0.001) and negatively vs. the ratio (-0.225/<0.021). 25-OH-VitD correlated negatively with the ratio (-0.757/<0.001). 1,25-DiOH-VitD had positive correlation vs. the ratio (0.592/<0.001).

For abortion cases, age correlated positively with gravidity (0.406/<0.02), parity (0.535/= 0.0024), BMI (0.757/<0.001), and pregnancy duration (0.337/= 0.046), and, negatively vs. 1,25-DiOH-VitD (-0.669/<0.001)and the ratio (-0.380/<0.028).Gravidity correlated positively with parity (0.815/<0.001) and duration of pregnancy (0.565/=

0.0013), and, negatively vs. each of 1,25-DiOH-VitD (-0.333/= 0.048) and the ratio (-0.381/ = 0.027). Parity related positively with duration of pregnancy (0.358/= 0.036). BMI associated negatively with 1,25-DiOH-VitD (-0.385/= 0.026). Pregnancy duration correlated negatively vs. 1.25-DiOH-VitD (-0.371/<0.031) and the ratio (-0.490/<0.006). 25-OH-VitD correlated negatively vs. the ratio (-0.773/<0.001). 1,25-DiOH-VitD correlated positively vs. the ratio (0.665/<0.001).

For the ectopic pregnancy cases, age correlated gravidity (0.884/<0.001), parity positively vs. (0.927 < 0.001), and (0.922/<0.001),BMI but. negatively vs. 1,25-DiOH-VitD (-0.639/<0.005) and the ratio (-0.527/<0.02). Gravidity correlated positively vs. parity (0.906/<0.001) and BMI (0.854/<0.001), and, negatively vs. 1,25-DiOH-VitD (-0.565/<0.013). Parity correlated positively with BMI (0.933/<0.001), and, negatively vs. 1,25-DiOH-VitD (-0.624/<0.006) and the ratio (-0.437/<0.05). BMI correlated positively vs. duration of pregnancy (0.433/<0.05), and, negatively vs. 1,25-DiOH-VitD (-0.443/= 0.044). Pregnancy duration related 1,25-DiOH-VitD (-0.504/<0.027). negatively vs. 25-OH-VitD correlated negatively vs. the ratio (-0.690/=0.002). 1,25-DiOH-VitD correlated positively with the ratio (0.647 / = 0.004).

Among PROMs cases, age correlated positively vs. gravidity (0.578/<0.018), parity (0.578/<0.018), BMI (0.727/= 0.002), 1,25-DiOH-VitD (0.500/<0.038) and the ratio (0.474/<0.05). Gravidity correlated positively vs. parity (1.0/<0.001), BMI (0.517/= 0.03) and duration of pregnancy (0.459/<0.05), but, negatively vs. 25-OH-VitD (-0.543/<0.026). Parity associated positively vs. BMI (0.517/= 0.03) and pregnancy duration (0.467/<0.05), but, negatively vs. (-0.543/<0.026). 25-OH-VitD BMI positively correlated with 1,25-DiOH-VitD (0.800/<0.001) and the ratio (0.655/= 0.007). 25-OH-VitD correlated positively with 1,25-DiOH-VitD (0.468/<0.048). 1,25-DiOH-VitD correlated positively vs. the ratio (0.786 < 0.001).

Within the preterm labor cases, age correlated positively vs. gravidity (0.898/<0.001), parity (0.908/<0.001), BMI (0.952/<0.001), and the ratio (0.452/<0.042), but, negatively vs. 25-OH-VitD (-0.439/<0.05). Gravidity correlated positively vs. parity (0.933/<0.001), BMI (0.802/<0.001) and the ratio (0.503/=0.025). Parity correlated positively vs. BMI (0.835/<0.001) and the ratio (0.454/=0.04), but, negatively with 25-OH-VitD (-0.454/= 0.04). BMI negatively with 25-OH-VitD associated (-0.500/<0.027), but, positively vs. the ratio (0.548/= 0.016). 1,25-DiOH-VitD correlated positively vs. the ratio (0.786/<0.001).

Discussion

Age was insignificantly different among our complicated groups. It had significant difference only when controls are compared to preeclamptics. Preeclampsia usually attacks extremely aged mothers; mainly primigravida. Gravidity showed a significant difference between the controls and GDM groups. Repeated pregnancy is a definite risk factor for GDM development. BMI was significantly different only comparing GDM and controls. GDM usually associates with higher BMI. As the gestational age is unique for the occurring complication (e.g., preeclampsia develops after 20 weeks of gestation, abortion frequently happens in the 1st trimester of pregnancy, PROMs occurs after 37th weeks and before onset of labor and GDM is common at beginning of the last trimester), it showed significant differences comparing all groups [15-19].

Considering the association between VitD deficiency and risk of feto-maternal pregnancy complications, the bending and intriguing questions are: How to achieve and specify the optimal pregnancy VitD level? What optimum biomarker and cutoffs to measure?, and, how to determine the real benefits of VitD supplementation in pregnancy? [20]. Even implementing a personal physiological response-dependent VitD signature turned confusing due to varying physiological responses at same supplementation dose and increment plasma 25-OH-VitD levels [21]. Reports related VitD level to clinical morbidities using serum 25-OH-VitD as a VitD biomarker - based on its higher stability and Such validity longer biological half-life. is substantiated by the significant correlation of serum 25-OH-VitD with non-calcemic clinical outcomes and its existence at high cellular levels. Both assumptions are not unquestionably correct [22, 23]. Indeed, the present study and a long list of previous studies, including ours, had shown that serum 25-OH-VitD does not differentiate apparently healthy controls from patients with different comorbidities including pregnancy, diabetes and infection [4, 8, 24-31]. This could be partially due to our and others observation of widely spreading hypovitaminosis D among population [4, 11]. 25-OH-VitD is produced from several sources and leak to serum without direct reflection of cellular contents [32, 33].

Serum level of 1,25-diOH-VitD was considered a less accurate predictor of intracellular VitD concentration. This came from the notion that 1,25-diOH-VitD intracellular level exceeds its serum level by several folds along with extra-renal sources for it [22]. Indeed, cells have concentrating modalities for lipophilic ligands. Moreover, serum 1,25-diOH-VitD is already VDR receptor saturating.

Therefore, the intracellular receptor supersaturating concentration is not required, albeit, for potential VDR-independent actions [34]. We resourced 1,25-diOH-VitD since several studies reported its superiority over 25-OH-VitD as an independent clinical correlate [31, 35-37]. 25-OH-VitD is the local precursor and increases in its circulating levels parallel consequent increases in 1,25-diOH-VitD however, their relationship is not always straightforward [30, 38]. For instance, adequate circulating 1,25-diOH-VitD levels are necessary for sufficient 25-OH-VitD availability in VitD target cells, e.g., monocytes [39]. Moreover, pregnant Brazilian women insufficient in VitD at baseline had higher increases in 1,25-diOH-VitD concentrations, over pregnancy time, compared to women with sufficient VitD levels [3]. VitD supplementation do not increase rate of sufficiency in all occasions [40]. However, 1,25-diOH-VitD levels had no seasonal variation like that of 25-OH-VitD in healthy Canadians [41]. a study found a strong correlation Moreover, between circulating 1,25-diOH-VitD and 25-OH-VitD levels throughout pregnancy and that 1,25-diOH-VitD levels at 12 weeks' gestation are approximately triple that of normal nonpregnant female [42].

Our controls were overwhelmingly VitD insufficient which is in accord with another Saudi study reporting 3.5% VitD sufficiency rate among pregnant women [43]. However, significantly lower 25-OH-VitD level vs. controls was recorded only for our GDM women among other complications. Ironically, those with ectopic pregnancy showed nonsignificant higher, while, other complications had nonsignificantly lower levels than controls. Grouping deficient and insufficient subjects revealed decreasing serum 25-OH-VitD rates as follows: Controls, ectopic pregnancy, abortion, GDM, preeclampsia, preterm labour, and PROMs. Oppositely, 1,25-diOH-VitD levels showed marked reductions in each of 6 complicated groups vs. controls. The complicated women groups were nonsignificantly different. Changes in 1,25-diOH-VitD/25-OH-VitD ratio mirrored 1,25-diOH-VitD pattern. While sensitivity and specificity of both of 1,25-diOH-VitD and the ratio were very high, those of 25-OH-VitD were very low in differentiating between our controls and complicated pregnancy. Pregnancy hypovitaminosis D in developing countries and its associated adverse feto-maternal hazards is a commonplace [10, 44]. Similar to our GDM women, studies had correlated hypovitaminosis D higher risk of development of GDM, miscarriage and still birth in the 2nd and 3rd trimester [23, 45-47]. Despite the wide spreading 25-OH-VitD deficiency in controls (71.2 vs. 83.3%), women with GDM had a 2.66-fold increased risk of

being deficient status [48].

Other studies conducted in Saudi Arabia, USA and China did not find association between adverse pregnancy outcomes and low 25-OH-VitD level except for high prevalence of abortions among other outcomes [8, 24, 26]. Surprisingly, the Chinese study found higher prevalence of GDM and preterm delivery among pregnant women with high 25-OH-VitD level despite older maternal age and higher BMI [18]. Actually 5% of women with GDM of our study had normal VitD levels. Rodriguez et al reported no association of VitD concentration and GDM, preterm delivery and IUGR [25]. Also, hypovitaminosis D in early pregnancy did not associate with adverse pregnancy outcomes [26]. GDM had no association with vitamin D deficiency [27, 28].

Similar to the trend of our results, higher risks of development of preeclampsia, GDM, preterm and cesarean delivery and IUGR were related to hypovitaminosis D [49, 50]. Normal 25-OH-VitD levels decrease the risk of preeclampsia and IUGR [51]. To establish a causal relationship, large VitD supplementation RCTs must guarantee optimum feto-maternal outcomes [52]. On the contrary, vitamin D supplementation did not affect the incidence of adverse pregnancy outcomes like preeclampsia, GDM and IUGR [53]. Conflictingly, supplementation with VitD in early pregnants diminished GDM incidence in pregnant with hypovitaminosis D [54]. Increased 25-OH-VitD levels after VitD supplementation correlate with significant reduction in rate of preterm labour, pre-eclampsia, and GDM in pregnant Indian women [43]. Women with persistent 25-OH-VitD deficiency up to 26th weeks of gestation had a 4.46-fold elevated risk for GDM [55]. Significant reduction in GDM risk was observed with increasing 25-OH-VitD levels [56].

Among our preeclampsia patients, 97% were VitD deficient/insufficient. Hypovitaminosis D is a risk factor for severe preeclampsia and season-dependent variation in incidence of preeclampsia seems to correlate with VitD level [57]. Low 25-OH-VitD levels and placental VitD receptor expression negatively correlated with preeclampsia [58]. 92% of our abortion patients had VitD deficiency/insufficiency. Relationship between low VitD levels and miscarriage stems from VitD regulation of genes concerned with implantation, trophoblastic invasion, angiogenesis, immunomodulation, suppression of inflammation and protection from infection [31, 59, 60]. 25-OH-VitD levels were lower in non-gravid women with history of pregnancy loss than normal non-gravid and pregnant women [61]. Reduced VitD levels among pregnant

women were correlated with increased production of proinflammatory cytokines [59]. Women with PROMs preterm labor were 100% VitD and deficient/insufficient in our study. Mothers with preterm labor are three times more likely to have insufficiency when compared to full-term mothers [62]. Hypovitaminosis D is associated with high prevalence of infections, particularly Gardnerella vaginalis; commonly encountered in cases of PROMs and preterm labor [63-65]. VitD deficiency increased susceptibility to microbial infection due to macrophage dysfunction with defect in toll-like mediated action of the antibacterial peptide cathelicidin [66].

BMI inversely correlates with 25-OH-VitD levels at med-gestation and postpartum [45]. Our controls had negative correlations contrasting 1,25-diOH-VitD and the ratio vs. pregnancy duration, BMI and 25-OH-VitD, while 25-OH-VitD had no such correlations. Among our preeclamptics, age, gravidity and BMI correlated negatively with 1,25-diOH-VitD. Our GDM group presented negative correlation between age vs. 1,25-diOH-VitD and the ratio, while gravidity and BMI had negative correlation with the ratio. Our abortion cases had negative correlation contrasting age, gravidity and BMI vs. 1,25-diOH-VitD and the ratio. 1,25-diOH-VitD negative correlation vs. age, gravidity, parity and BMI among our ectopics. In our PROMs cases, age and BMI had positive correlation vs. 1,25-diOH-VitD and the ratio, while, gravidity and parity correlated negatively vs. 25-OH-VitD. Our preterm labor cases showed positive correlation between age, gravidity, parity and BMI vs. the ratio but a negative correlation between age, parity and BMI vs. 25-OH-VitD. Reportedly, gestational age correlates positively with 25-OH-VitD levels and had stronger association with 1,25-diOH-VitD [3]. Disagreeable, serum 25-OH-VitD level neither increased the risk of preterm birth nor correlated with gestational age or BMI [67]. A big supplementation study VitD conducted in Bangladesh, with a widespread VitD deficiency and fetal-infant growth restriction, showed that supplementation from mid-pregnancy to delivery or 6 months postpartum had no significant effect on pregnancy feto-maternal clinical outcomes, despite normalizing 25-OH-VitD levels [68].

Our study is one of the few studies which evaluated the association of variations in 1,25-diOH-VitD and its ratio to 25-OH-VitD with pregnancy complications. Physiologically, 1,25-diOH-Vit-D levels increase with gestational age to reach a maximum in 3rd trimester without increases in 25-OH-VitD levels [30]. In our study, while 25-OH-VitD correlated negatively only with GDM, 1.25-diOH-VitD and its ratio to 25-OH-Vit-D correlated negatively with all of pregnancy complications investigated. This may help clearing the discrepancy among the previous studies utilizing 25-OH-Vit-D as the sole VitD biomarker for adverse pregnancy complications. We claimed that addition of 1,25-diOH-VitD and the ratio in evaluating implication in pregnancy complications are clinically meaningful than 25-OH-VitD alone. They both showed higher sensitivity and specificity in differentiating healthy controls vs. complicated pregnancies. However, bigger multi-centric longitudinal and supplementation RCTs are strongly warranted to validate and generalize these assumptions. Several clinical outcomes, including infection, kidney function, diabetes and inflammation, were inversely correlated with circulating 1,25-diOH-VitD levels but not 25-OH-VitD [29, 31, 37].

The norms and biomarkers for VitD levels have to be reevaluated, particularly for pregnancy and its complication, owing to the actual heterogeneity of the world climate and populations. Major potential causes of the widely spreading VitD deficiency we observed include: 1) the traditionally low VitD foods, 2) a scalding local summer sun and ice-cold winter that give very narrow room for sun exposure, 3) low supplement intake, and, 4) absence of a national mandatory VitD food fortification program.

The difficulties faced us carrying out this study included: 1) Absence of a reasonable number of other pregnancy complications such as congenital anomalies, stillbirth, isolated IUGR, 2) we could not correlate findings with comorbid rate of infection and inflammation, length of hospital stays, and long-term outcomes, & 3) Longitudinal analysis was not accessible.

VitD deficiency/insufficiency widely is spreading in the studied population of pregnant women. Levels of 25-OH-VitD were not different among controls and complicated pregnancies except for a significantly lower level in GDM. 25-OH-VitD levels did not correlate with most of the characteristics of participants and pregnancy complications. 1,25-diOH-VitD and its ratio to 25-OH-VitD revealed very highly significant reduction in complicated pregnancy and were highly sensitive and specific in controls differentiating from complicated pregnancies. They also correlate with most of the of the participating characteristics women. 1,25-diOH-VitD and its ratio to 25-OH-VitD are potentially more functionally relevant biomarkers for VitD status particularly during pregnancy.

Acknowledgments

We highly appreciate the generous support from

Dr Atef abdelmageed (Head, Obstetrics and Gynecology Department) and Mr. Mohammed Fahman (Manager of the Laboratory, Maternity and Children Hospital, Sakaka, Aljouf, Saudi Arabia) for facilitating the collection of the samples and profiling our patients.

Other than clinical profiling of the patients and collecting samples by IAB, all authors contributed equally to this article.

Funding

This study was generously funded by the Deanship for Research, Jouf University, Sakaka, Saudi Arabia (#39/950).

Competing Interests

The authors have declared that no competing interest exists.

References

- Gallagher JC, Sai AJ. Vitamin D insufficiency, deficiency, and bone health. J Clin Endocrinol Metab. 2010; 95: 2630-3. doi: 10.1210/jc.2010-0918.
- 2. Carlberg C. Vitamin D: A micronutrient regulating genes. Curr Pharm Des. 2019; 25(15): 1740-6. doi: 10.2174/1381612825666190705193227.
- Figueiredo ACC, Cocate PG, Adegboye ARA, et al. Changes in plasma concentrations of 25-hydroxyvitamin D and 1,25-dihydroxyvitamin D during pregnancy: A Brazilian cohort. Eur J Nutr. 2018; 57(3): 1059-72. doi: 10.1007/s00394-017-1389-z.
- Abdulrahman A. Alduraywish, Abdullah Z. Almani, et al. Vitamin D insufficiency in type 2 diabetic patients correlates the disease prognostic indices: A northern Saudi status. International Medical J. 2019; 26(4): 303-10.
- Holick MF, Binkley NC, Bischoff-Ferrari HA, et al. Evaluation, treatment, and prevention of vitamin D deficiency: An endocrine society clinical practice guideline. J Clin Endocrinol Metab. 2011; 96: 1911-30. doi: 10.1210/jc.2011-0385.
- [Internet] The Scientific Advisory Committee on Nutrition recommendations on vitamin D. https://www.gov.uk/government/ publications/sacn-vitamin-d-and-health-report. (Accessed December 25, 2019).
- Dawodu A, Wagner CL. Mother-child vitamin D deficiency: An international perspective. Arch Dis Child. 2007; 92: 737-40. doi: 10.1136/adc.2007.122689.
- Al-Shaikh GK, Ibrahim GH, Fayed AA, et al. Impact of vitamin D deficiency on maternal and birth outcomes in the Saudi population: A cross-sectional study. BMC Pregnancy Childbirth. 2016; 16(119):1-9. doi: 10.1186/s12884-016-0901-4.
- 9. Wagner CL, Hollis BW. The implications of vitamin D status during pregnancy on mother and her developing child. Front Endocrinol (Lausanne). 2018; 9: 500. doi: 10.3389/fendo.2018.00500.
- De-Regil LM, Palacios C, Lombardo LK, et al. Vitamin D supplementation for women during pregnancy. Sao Paulo Med J. 2016; 134(3): 274-5. doi: 10.1590/1516-3180.20161343T2.
- van der Pligt P, Willcox J, Szymlek-Gay EA, et al. Associations of maternal vitamin D deficiency with pregnancy and neonatal complications in developing countries: A systematic review. Nutrients. 2018; 10(5): 640;1-22. doi: 10.3390/nu10050640.
- 12. Katrinaki M, Kampa M, Margioris A, et al. Vitamin D levels in a large Mediterranean cohort: reconsidering normal cut-off values. Hormones (Athens). 2016; 5(2): 205-23. doi: 10.14310/horm.2002.1674.
- 13. [Internet] American Association of Clinical Endocrinologists. https://www.aace.com/sites/default/files/2019-02/Vitamin_D_Defici ency_formatted.pdf (Last accessed December 28, 2019).
- 14. [Internet] The Office of Dietary Supplements, NIH. https://ods.od.nih.gov/factsheets/VitaminD-HealthProfessional/ (Last accessed November 30, 2019).
- Graham A, Devarajan S, Datta S. Complications in early pregnancy. Obstetrics, Gynaecology and Reproductive Medicine. 2015; 25(1): 1-5. doi:https://doi.org/10.1016/j.ogrm.2014.10.009

- Neiger R. Long-term effects of pregnancy complications on maternal health: A review. J Clin Med. 2017; 6(8): 76. doi: 10.3390/jcm6080076.
- Alves NCC, Feitosa KMA, Mendes MES, Caminha MFC. Complications in pregnancy in women aged 35 or older. Rev Gaucha Enferm. 2017; 38(4e): 2017-42. doi: 10.1590/1983-1447.2017.04.2017-0042.
- [Internet] womenshealth.gov. Pregnancy complications. https://www.womenshealth.gov/pregnancy/youre-pregnant-now-wh at/pregnancy-complications.
- [Internet] WHO. Managing complications in pregnancy and childbirth: A guide for midwives and doctors. 2nd Edition, 2017. https://apps.who.int/iris/bitstream/handle/10665/255760/978924156 5493-eng.pdf?sequence=1.
- Ciebiera M, Wojtyła C, Łukaszuk K, et al. The role of vitamin D in perinatology - What do we know at the beginning of 2018? Arch Med Sci. 2019; p:1-14 (ahead of print). doi: https://doi.org/10.5114/ aoms.2019.81747.
- Shirvani A, Kalajian TA, Song A, et al. Variable genomic and metabolomic responses to varying doses of vitamin D supplementation. Anticancer Res. 2020; 40(1): 535-43. doi: 10.21873/anticanres.13982.
- Jones G. Metabolism and biomarkers of vitamin D. Scandinavian J. Clinical & Laboratory Investigation. 2012; 243: 7-13. doi: 10.3109/00365513.2012.681892.
- 23. Wen J, Hong Q, Zhu L, et al. Association of maternal serum 25-hydroxyvitamin D concentrations in second and third trimester with risk of gestational diabetes and other pregnancy outcomes. Int J Obes (Lond). 2017; 41(4): 489-96. doi: 10.1038/ijo.2016.227.
- Zhou J, Su L, Liu M, et al. Associations between 25-hydroxyvitamin D levels and pregnancy outcomes: a prospective observational study in southern China. Eur J Clin Nutr. 2014; 68(8): 925-30. doi: 10.1038/ejcn.2014.99.
- Rodriguez A, Garcia-Esteban R, Basterretxea M, et al. Associations of maternal circulating 25-hydroxyvitamin D3 concentration with pregnancy and birth outcomes. BJOG. 2015; 122: 1695-704. doi: 10.1111/1471-0528.13074.
- Flood-Nichols SK, Tinnemore D, Huang RR, et al. Vitamin D deficiency in early pregnancy. PLoS One. 2015; 10: 1-15. doi: 10.1371/journal.pone.0123763.
- Baker AM, Haeri S, Camargo Jr CA, et al. First-trimester maternal vitamin D status and risk for gestational diabetes (GDM) a nested case-control study. Diabetes Metab Res Rev. 2012; 28: 164-8. doi: 10.1002/dmrr.1282.
- Maklouba M, Nelson SM, Savvidou M, et al. First-trimester circulating 25-hydroxyvitamin D levels and development of gestational diabetes mellitus. Diabetes Care. 2011; 34: 1091-3. doi: 10.2337/dc10-2264.
- Zittermann A, Schleithoff SS, Götting C, et al. Calcitriol deficiency and 1year mortality in cardiac transplant recipients. Transplantation. 2009; 87: 118-24. doi: 10.1097/TP.0b013e31818c2708.
- Zittermann A, Kuhn J, Ernst JB, et al. 25-hydroxyvitamin D, 1,25-dihydroxyvitamin D and postoperative outcome in cardiac surgery. J Clin Endocrinol Metab. 2015; 100: 72-80. doi: 10.1210/jc.2014-3013.
- Zittermann A, Kuhn J, Ernst JB, et al. Circulating 25-hydroxyvitamin D and 1,25-dihydroxyvitamin D concentrations and postoperative infections in cardiac surgical patients: The CALCITOP-Study. PLoS One. 2016; 11(6e): 1-11. doi: 10.1371/journal.pone.0158532.
- Albert PJ, Proal AD, Marshall TG. Vitamin D: The alternative hypothesis. Autoimmun Rev. 2009; 8(8): 639-44. doi: 10.1016/j.autrev. 2009.02.011.
- Carlberg C, Haq A. The concept of the personal vitamin D response index. J Steroid Biochem Mol Biol. 2018; 175: 12-7. doi: 10.1016/j.jsbmb.2016.12.011.
- Balasuriya CND, Larose TL, Mosti MP, et al. Maternal serum retinol, 25(OH)D and 1,25(OH)₂D concentrations during pregnancy and peak bone mass and trabecular bone score in adult offspring at 26-year follow-up. PLoS One. 2019; 14(9e): 1-15. doi: 10.1371/journal.pone. 0222712.
- Blaney GP, Albert PJ, Proal AD. Vitamin D metabolites as clinical markers in autoimmune and chronic disease. Ann N Y Acad Sci. 2009; 1173: 384-90. doi: 10.1111/j.1749-6632.2009.04875.x.
- Brannon PM, Picciano MF. Vitamin D in pregnancy and lactation in humans. Annu Rev Nutr. 2011; 31: 89-115. doi: 10.1146/annurev.nutr.012809.104807.
- Ney J, Heyland DK, Amrein K, et al. The relevance of 25-hydroxyvitamin D and 1,25-dihydroxyvitamin D concentration for postoperative infections and postoperative organ dysfunctions in cardiac surgery patients: The eVIDenCe study. Clin Nutr. 2019; 38(6): 2756-62. doi: 10.1016/j.clnu.2018.11.033.
- Dini C, Bianchi A. The potential role of vitamin D for prevention and treatment of tuberculosis and infectious diseases. Ann Ist Super Sanita. 2012; 48: 319-27. doi: 10.4415/ANN_12_03_13.

- Gallieni M, Kamimura S, Ahmed A, et al. Kinetics of monocyte 1 alpha-hydroxylase in renal failure. Am J Physiol. 1995; 268(F): 746-53. doi: 10.1152/ajprenal.1995.268.4.F746.
- Awker AL, Herbranson AT, Rhee TG, et al. Impact of a vitamin D protocol in pregnancy at an urban women's health clinic. Ann Pharmacother. 2016; 50(11): 935-41. doi: 10.1177/1060028016657555.
- Rucker D, Allan JA, Fick GH, et al. Vitamin D insufficiency in a population of healthy western Canadians. CMAJ. 2002; 166(12): 1517-24. PMID: 12074117. PMCID: PMC113796
- Hollis BW, Johnson D, Hulsey TC, et al. Vitamin D supplementation during pregnancy: double-blind, randomized clinical trial of safety and effectiveness. J Bone Miner Res. 2011; 26(10): 2341-57. doi: 10.1002/jbmr.463.
- Al-Ajlan A, Krishnaswamy S, Alokail MS, et al. Vitamin D deficiency and dyslipidemia in early pregnancy. BMC Pregnancy Childbirth. 2015; 15(314):1-8. doi: 10.1186/s12884-015-0751-5.
- 44. Sablok A, Batra A, Thariani K, et al. Supplementation of vitamin D in pregnancy and its correlation with feto-maternal outcome. Clin Endocrinol (Oxf). 2015; 83(4): 536-41. doi: 10.1111/cen.12751.
- Guariguata L, Linnenkamp U, Beagley J, et al. Global estimates of the prevalence of hyperglycaemia in pregnancy. Diabetes Res Clin Pract. 2014; 103: 176-85. doi: 10.1016/j.diabres.2013.11.003.
- Pleskačová A, Bartáková V, Pácal L, et al. Vitamin D status in women with gestational diabetes mellitus during pregnancy and postpartum. Biomed Res Int. 2015; 260624: 1-7. doi: 10.1155/ 2015/260624.
- Al-Ajlan A, Al-Musharaf S, Fouda MA, et al. Lower vitamin D levels in Saudi pregnant women are associated with higher risk of developing GDM. BMC Pregnancy Childbirth. 2018; 18(1): 1-7. doi: 10.1186/s12884-018-1723-3.
- Soheilykhah S, Mojibian M, Rashidi M, et al. Maternal vitamin D status in gestational diabetes mellitus. Nutr Clin Pract. 2010; 25(5): 524-7. doi: 10.1177/0884533610379851.
- Wei SQ, Qi HP, Luo ZC, et al. Maternal vitamin D status and adverse pregnancy outcomes: a systematic review and meta-analysis. J Matern Fetal Neonatal Med. 2013; 26(9): 889-99. doi: 10.3109/14767058. 2013.765849.
- Wei SQ. Vitamin D and pregnancy outcomes. Curr Opin Obstet Gynecol. 2014; 26(6): 438-47. doi: 10.1097/GCO.00000000000117.
- 51. Mairead E, Joy Y, Michael K, et al. Vitamin D status is associated with uteroplacental dysfunction indicated by pre-eclampsia and small-for-gestational-age birth in a large prospective pregnancy cohort in Ireland with low vitamin D status. Am J Clin Nutr. 2016; 104: 354-61. doi: 10.3945/ajcn.116.130419.
- 52. Sharma N, Nath C, Mohammad J. Vitamin D status in pregnant women visiting a tertiary care center of North Eastern India. J Family Med Prim Care. 2019; 8(2): 356-60. doi: 10.4103/jfmpc.jfmpc_404_18.
- Perez-Lopez FR, Pasupuleti V, Mezones-Holguin E, et al. Effect of vitamin D supplementation during pregnancy on maternal and neonatal outcomes: a systematic review and meta-analysis of randomized controlled trials. Fertil Steril. 2015; 103: 1278-88. doi: 10.1016/j.fertnstert.2015.02.019.
- Yap C, Cheung NW, Gunton JE, et al. Vitamin D supplementation and the effects on glucose metabolism during pregnancy: A randomized controlled trial. Diabetes Care. 2014; 37: 1837-44. doi: 10.2337/dc14-0155.
- 55. Xia J, Song Y, Rawal S, et al. Vitamin D status during pregnancy and the risk of gestational diabetes mellitus: A longitudinal study in a multiracial cohort. Diabetes Obes Metab. 2019; 21(8): 1895-905. doi: 10.1111/dom.13748.
- 56. Yin WJ, Tao RX, Hu HL, et al. The association of vitamin D status and supplementation during pregnancy with gestational diabetes mellitus: A Chinese prospective birth cohort study. Am J Clin Nutr. 2020; 111(1): 122-30. doi: 10.1093/ajcn/nqz260.
- Bodnar LM, Simhan HN, Catov JM, et al. Maternal vitamin D status and the risk of mild and severe preeclampsia. Epidemiology. 2014; 25: 207-14. doi: 10.1097/EDE.00000000000039.
- Hutabarat M, Wibowo N, Obermayer-Pietsch B, et al. Impact of vitamin D and vitamin D receptor on the trophoblast survival capacity in preeclampsia. PLoS One. 2018; 13(11): 1-16. e0206725. doi: 10.1371/journal.pone.0206725.
- Diaz L, Noyola-Martinez N, Barrera D, et al. Calcitriol inhibits TNF-alpha-induced inflammatory cytokines in human trophoblasts. J Reprod Immunol. 2009; 81: 17-24. doi: 10.1016/j.jri.2009.02.005.
- Börgermann J, Lazouski K, Kuhn J, et al. 1,25-Dihydroxyvitamin D fluctuations in cardiac surgery are related to age and clinical outcome. Crit Care Med. 2012; 40: 2073-81. doi: 10.1097/CCM.0b013e31824e8c42.
- Hou W, Yan XT, Bai CM, et al. Decreased serum vitamin D levels in early spontaneous pregnancy loss. Eur J Clin Nutr. 2016; 70(9): 1004-8. doi: 10.1038/ejcn.2016.83.

- Kassai MS, Cafeo FR, Affonso-Kaufman FA, et al. Vitamin D plasma concentrations in pregnant women and their preterm newborns. BMC Pregnancy Childbirth. 2018; 18(1): 1-8. doi: 10.1186/s12884-018-2045-1.
- Liu NQ, Hewison M. Vitamin D, the placenta and pregnancy. Arch Biochem Biophys. 2012; 523: 37-47. doi: 10.1016/j.abb.2011.11.018.
- Hensel KJ, Randis TM, Gelber SE, et al. Pregnancy-specific association of vitamin D deficiency and bacterial vaginosis. Am J Obstet Gynecol. 2011; 204(41e): 1-9. doi: 10.1016/j.ajog.2010.08.013.
- Haghdoost S, Pazandeh F, Darvish S, et al. Association of serum vitamin D levels and urinary tract infection in pregnant women: A case control study. Eur J Obstet Gynecol Reprod Biol. 2019; 243: 51-6. doi: 10.1016/j.ejogrb.2019.10.015.
- Dovnik A, Mujezinović F, Treiber M, et al. Seasonal variations of vitamin D concentrations in pregnant women and neonates in Slovenia. Eur J Obstet Gynecol Reprod Biol. 2014; 181: 6-9. doi: 10.1016/j.ejogrb. 2014.07.019.
- Yang L, Pan S, Zhou Y, et al. The correlation between serum vitamin d deficiency and preterm birth. Med Sci Monit. 2016; 22: 4401-5. doi: 10.12659/msm.898117.
- Roth DE, Morris SK, Zlotkin S, et al. Vitamin D supplementation in pregnancy and lactation and infant growth. N Engl J Med. 2018; 379(6): 535-46. doi: 10.1056/NEJMoa1800927.